

INTRODUCTION TO MOLECULAR MACHINERY

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Optimization At a Small Scale UCSD July 20-21

... and collaborators

Alex Ciudad”*A unified phenomenological analysis of the experimental data velocity curves in molecular motors*”

J. Chem. Phys. **128**, 225107(2008)

Alex Gómez-Marín... “*Two-state flashing molecular pumps*”,

Europhysics Letters **86**, 40002(2009)

Javier G. Orlandi...”*Theoretical study of a membrane channel gated by ATP*”,

Eur. Phys. J. E, DOI 10.1140/epje/i2009-10483-9 (2009)

Rubén Pérez... “*Theoretical analysis of the F1-ATPase experimental data*”

Preprint, 2009.....

Slides from:

“*Physical Biology of the Cell*” , R.Phillips et al... Garland (2009)

OUTLINE

1. INTRODUCTION
2. MOTORS
3. CHANNELS
4. PUMPS
5. CONCLUSSIONS

MOLECULAR MACHINERY

MOTORS: | LINEAR: kinesin
| ROTATORY: BFM, Fo, F1..

TRANSPORTERS: | PUMPS: Fo, Na-K ATPase
| CHANNELS: KscA, Cl(CFTR),..

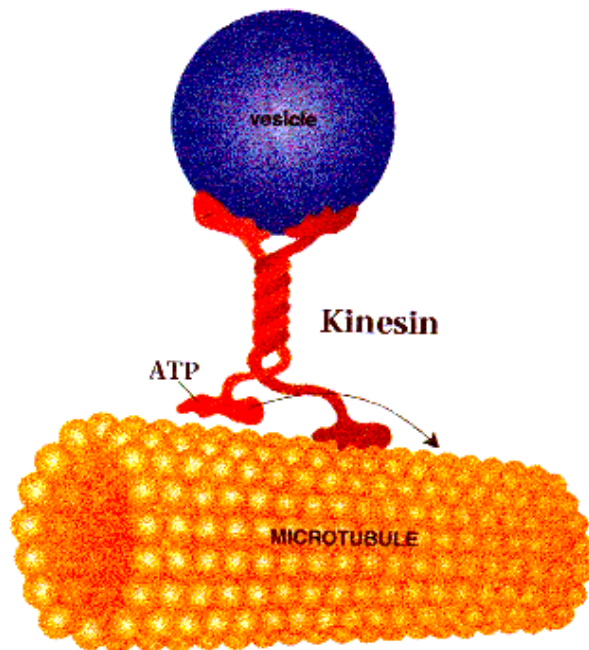
... and more: translocators, Polymerases, Ribosomes,...

and also artificial devices ...

EXAMPLES

LINEAR MOTORS

Kinesin

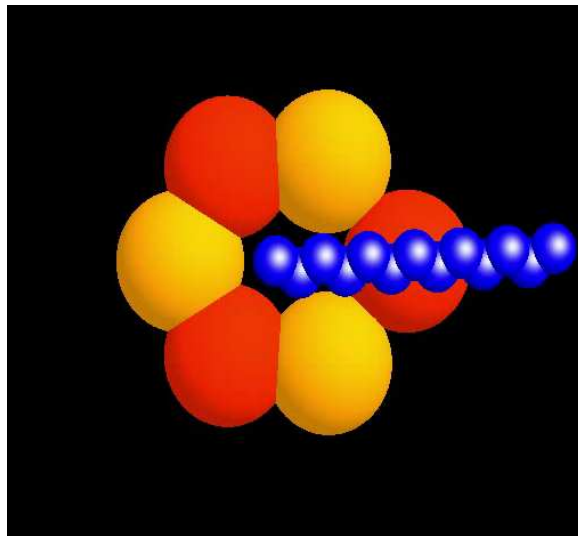


Maglev

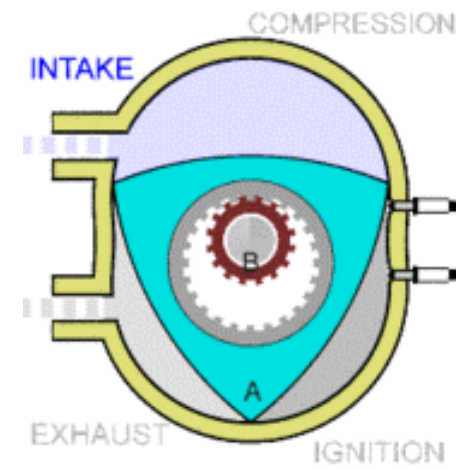


ROTATORY MOTORS

F1-ATPase



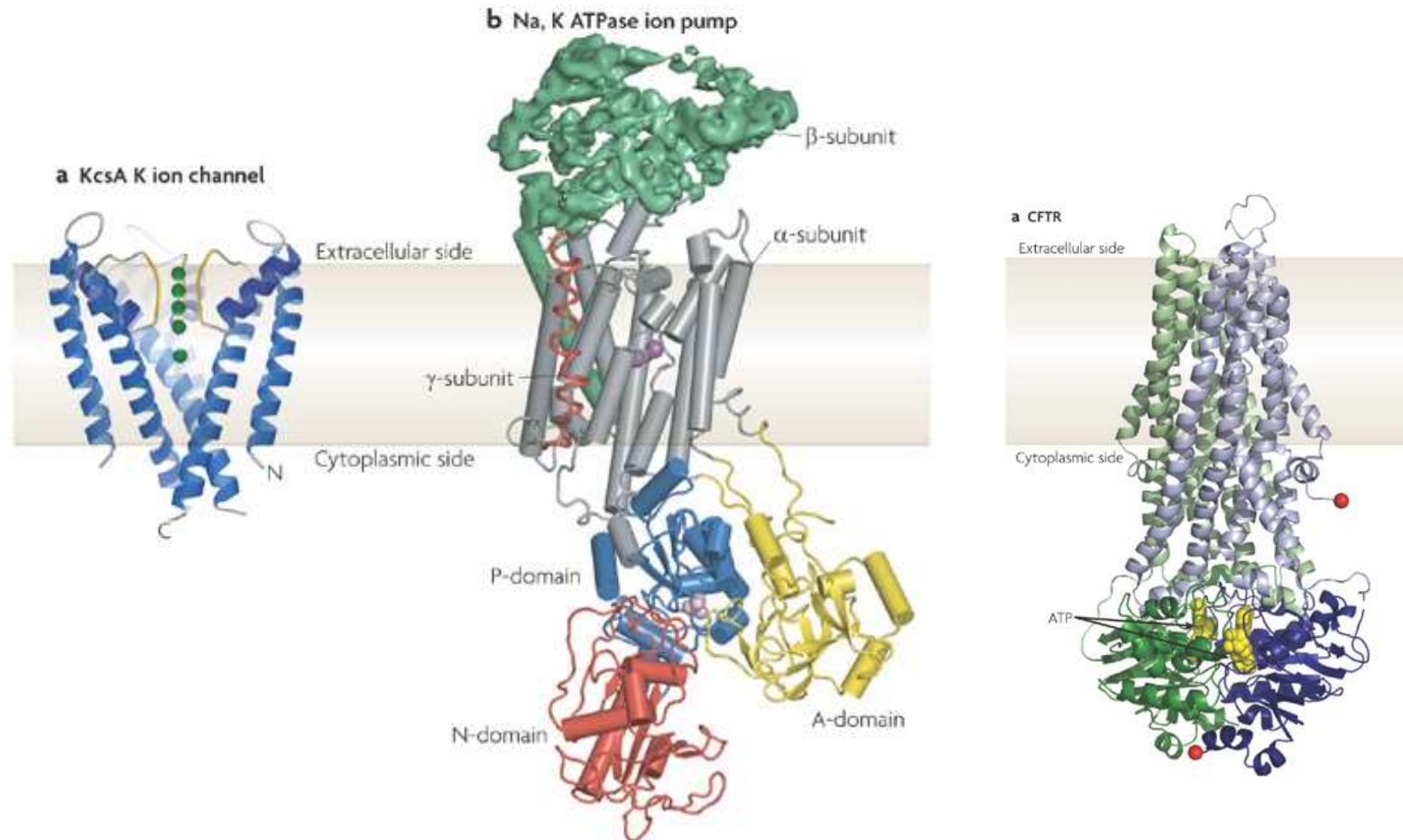
Wankel



K channel

Na-K ATP pump

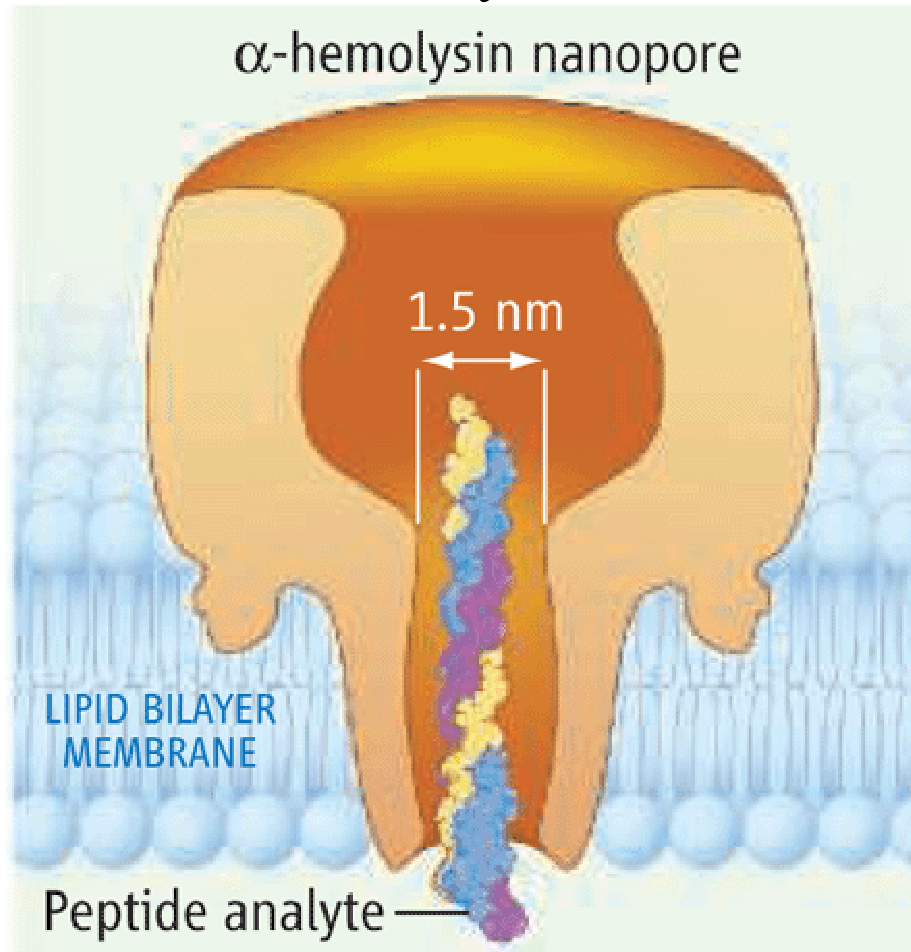
Cl (CFTR) ATP channel



From D.C. Gadsby, Nature Reviews-MCB, 10, 344 (2009)

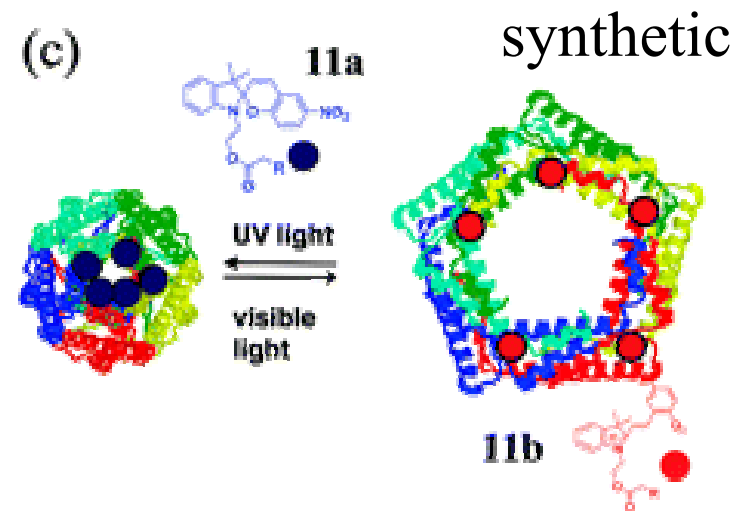
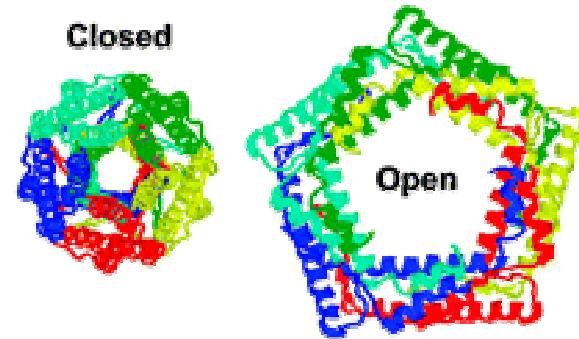
Channels

selectivity
 α -hemolysin nanopore



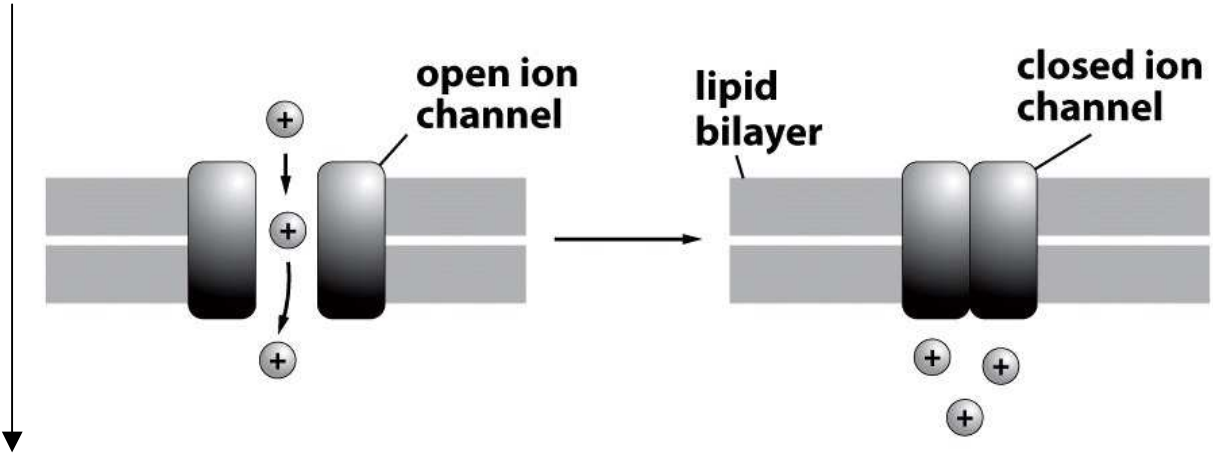
Martin & Siwy, *SCIENCE* **317**,3313(2007)

(a) MscL native



Feringa, *JOC Perspective* **72**, 6635 (2007)

Channel



Pump

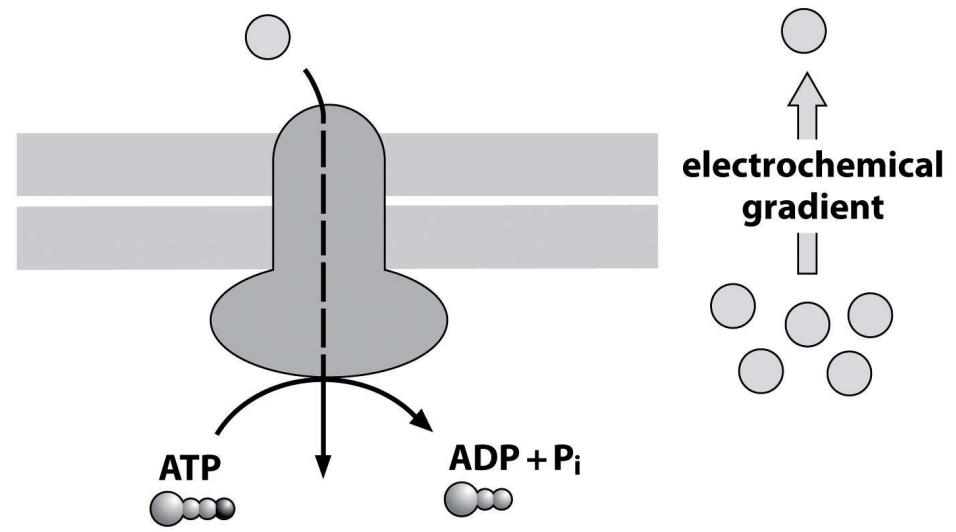
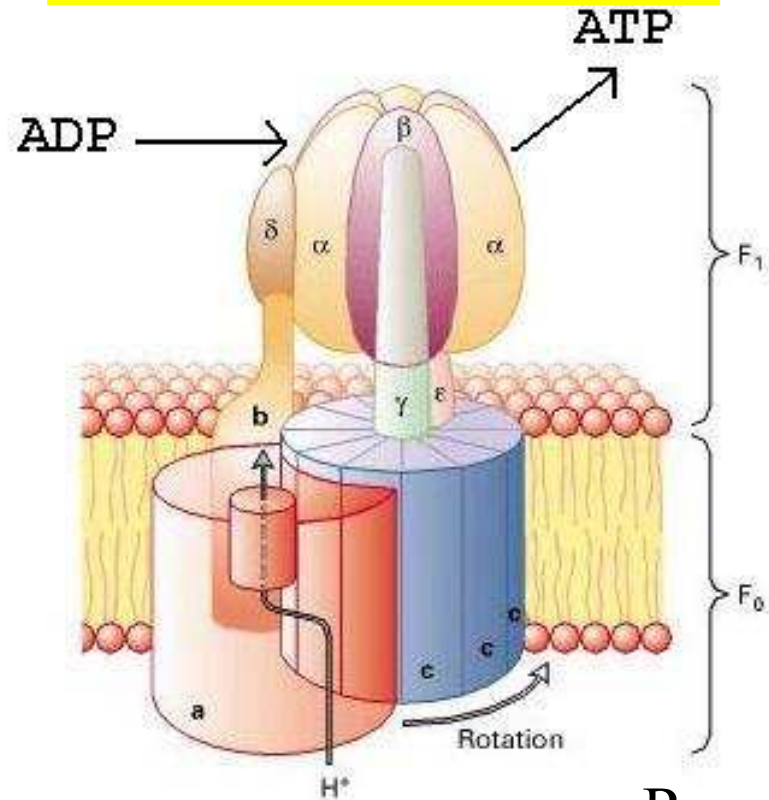


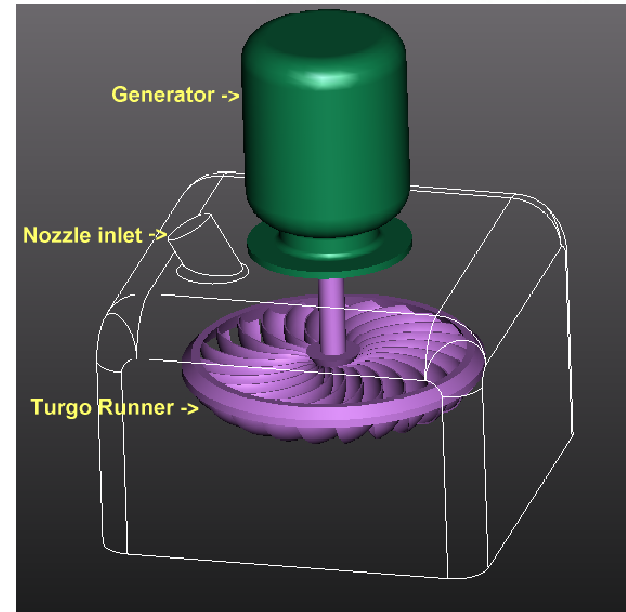
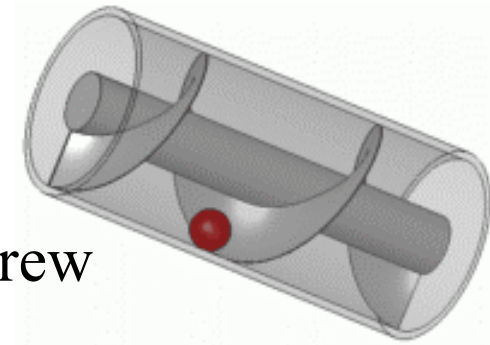
Figure 3.27 Physical Biology of the Cell (© Garland Science 2009)

MIXED MOTOR AND PUMP

F₀F₁-ATP Synthase



Archimedes screw



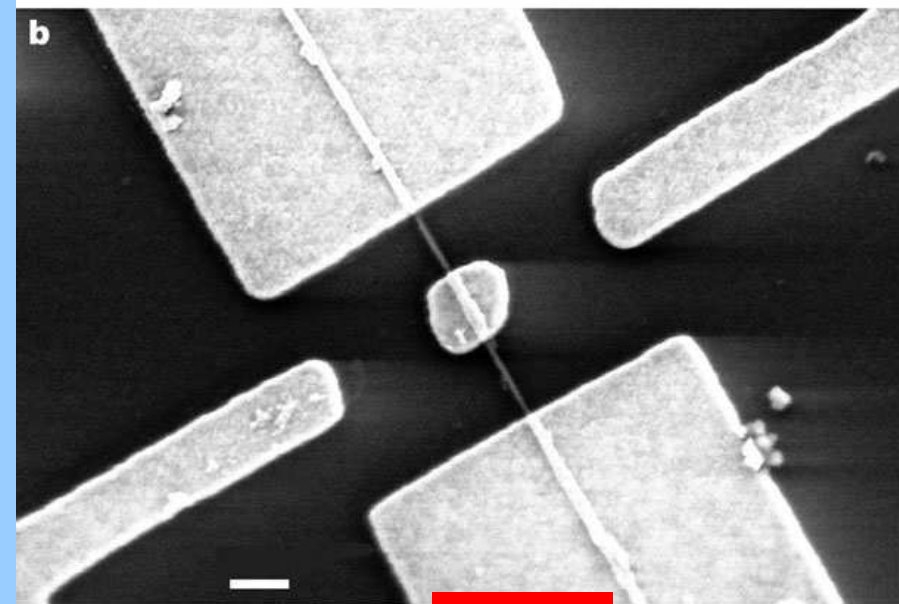
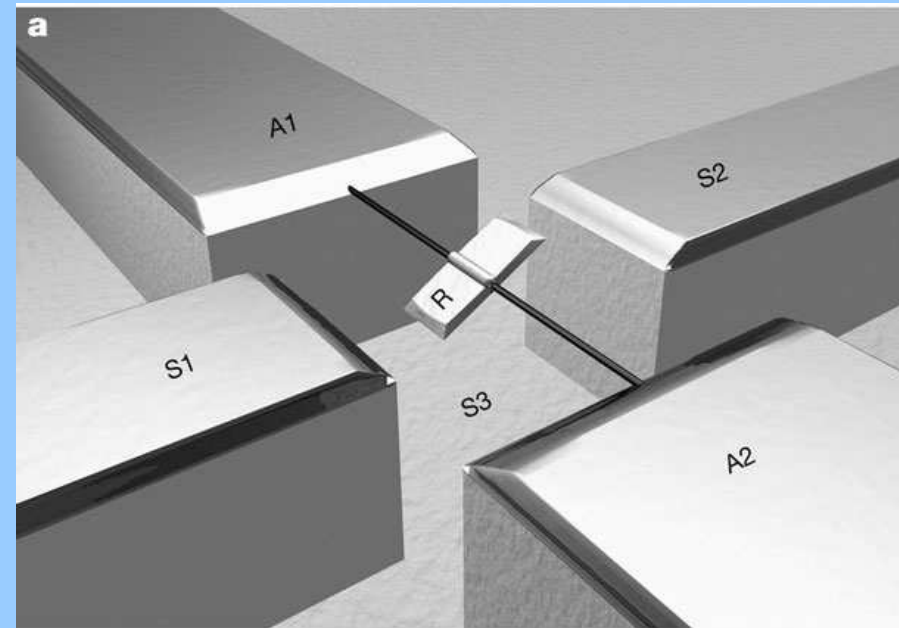
Reversible engine

ATP hydrolysis \rightarrow proton pump

ATP synthesis \leftarrow proton flux

Nano-devices

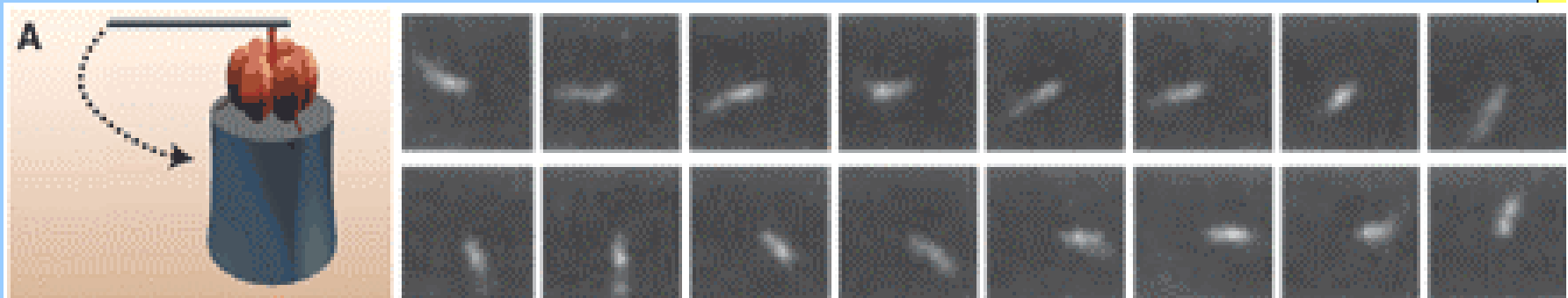
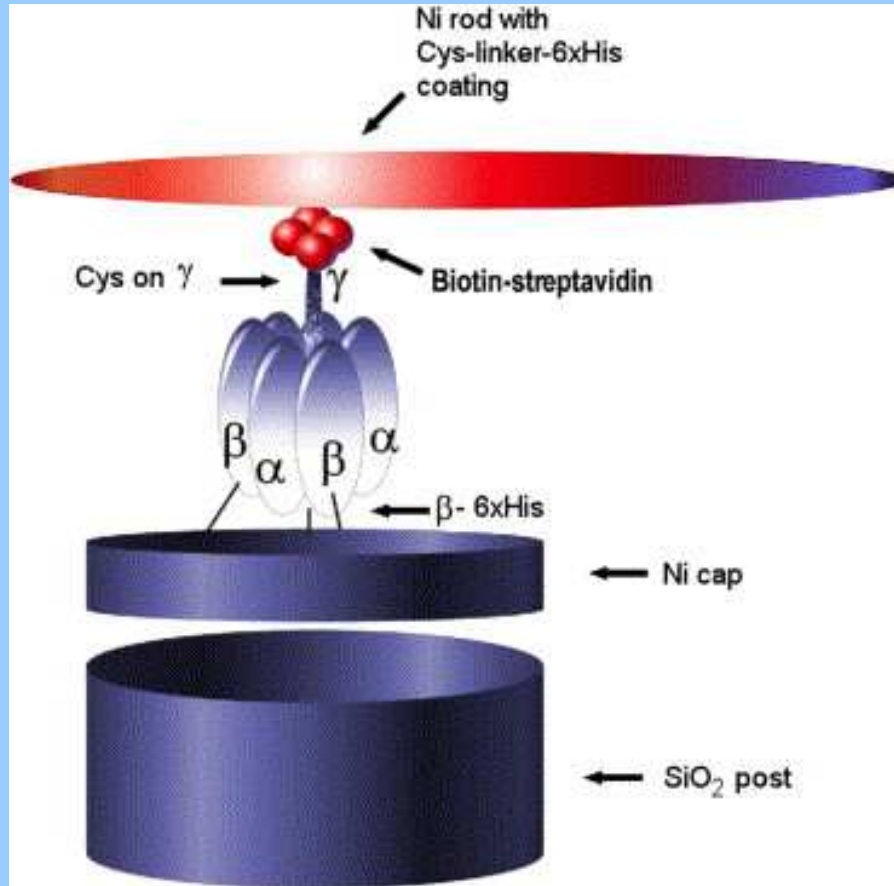
Fennimore, et al.
Nature **424**, 408 (2003)



[video](#)

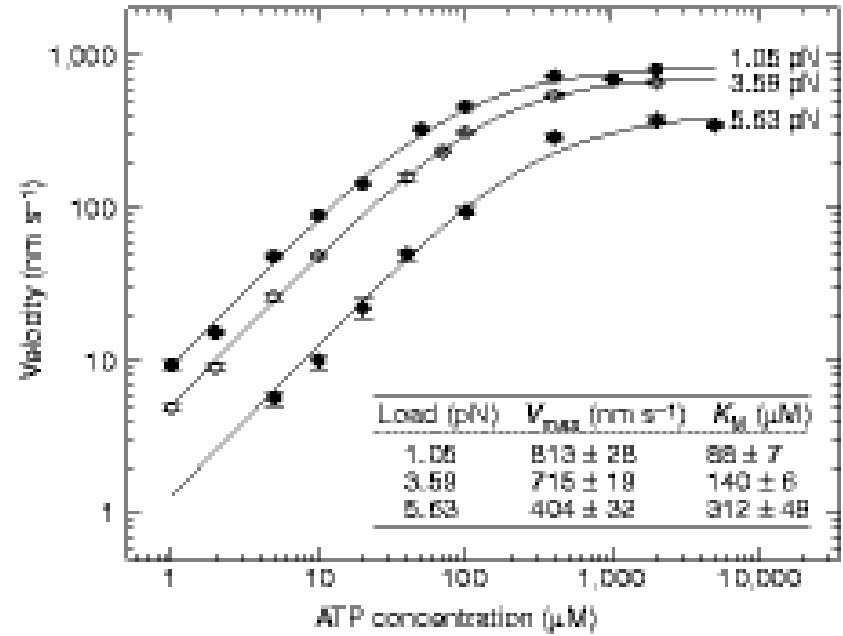
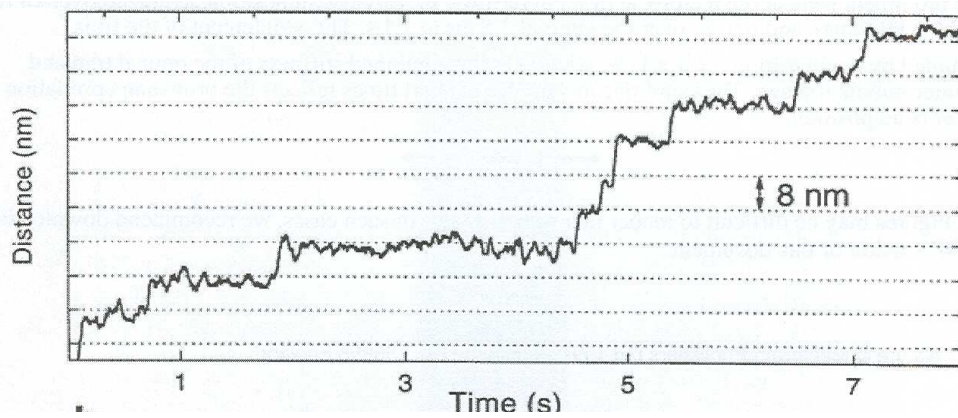
Mixed nanomotors: Rotatory. F1-ATPase

Song et al.,
Science **290**, 1555 (2000)

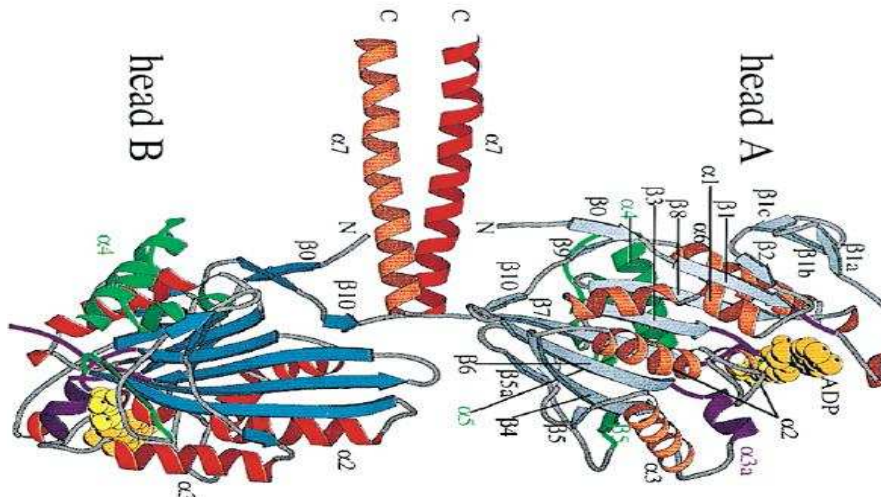


Levels of analysis

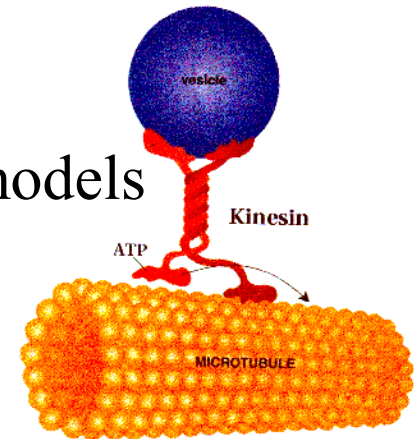
Experimental data: trajectories and mean observables



Structure and function



Mesoscopic models



Aim: A theoretical scenario for experimental data

Input or
control parameters:

[ATP] concentration

ATP-energy

Conservative forces: $F...$

Particle densities

Electrochemical potential

Friction

Outputs or observables:

Trajectories

Spatial steps

$\langle v \rangle$

Randomnes

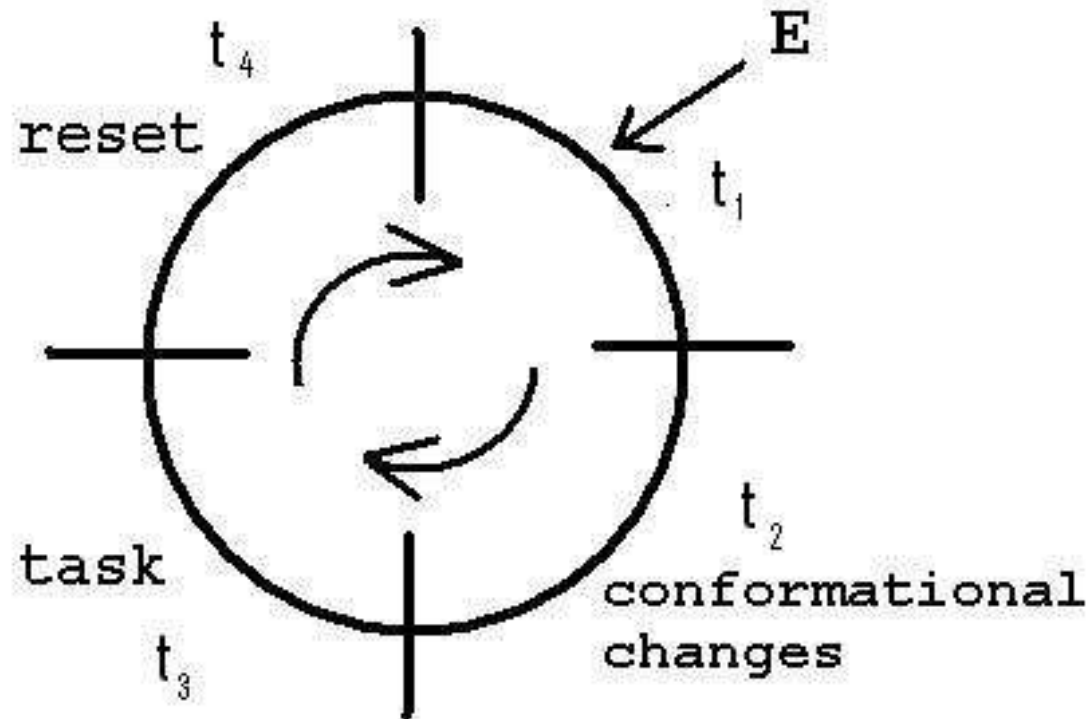
Power

Efficiency

$$v([ATP], F)$$

$$J([ATP], \Delta\phi, \rho_{in}, \rho_{out})$$

The starting point: the CYCLE



$$v = \frac{L_0}{t_1 + t_2 + \dots + t_n}$$

Physical and chemical elements

$$E_{in} \left| \begin{array}{l} \Delta G_{ATP} \sim 20k_B T \sim 90 \text{ pN.nm} \sim 51 \text{ meV} \\ smf = -\Delta\phi + k_B T \ln \frac{[Na]_{in}}{[Na]_{ex}} \end{array} \right.$$

Motive force $F_m = \frac{E_{in}}{L_0}$

Conservative force F_{ex}

Times: chemical (waiting and internal) and mechanical times.

Internal time $\rightarrow t_0$ Waiting time $\rightarrow t_0 \frac{k}{[ATP]}$

M-M

MOTORS

Scales: $\Delta G_{ATP} \sim 90 \text{ pN}\cdot\text{nm} \rightarrow F \sim \text{pN}; L_0 \sim \text{nm}$

Mechanical time

$$(\gamma_0 + \gamma_L) \frac{L_0}{t_{mech}} = F_m - F_{ex}$$

Analytical formula for motors

$$v = \frac{L_0}{t_0 \left(1 + \frac{k}{[ATP]} \right) + \frac{L_0(\gamma_0 + \gamma_L)}{F_m - F_{ex}}}$$

Too many free parameters!

Kinesin: $t_0 = 0.1 \text{ ms}$ $L_0 = 8 \text{ nm} \rightarrow v \sim 800 \text{ nm/s}$

CHANNELS

FLUX : diffusion down the gradient
driven by the membrane potential

GATING: open and close.
ATP hydrolysis
V membrane potential
density (M-M)
others

$$J = P_{open} * J_{free}$$

FLUX: Free open channel

$$\frac{\partial \rho}{\partial t} = D \frac{\partial^2 \rho}{\partial x^2}, \quad \rho(0) = \rho_0, \quad \rho(L) = \rho_1$$

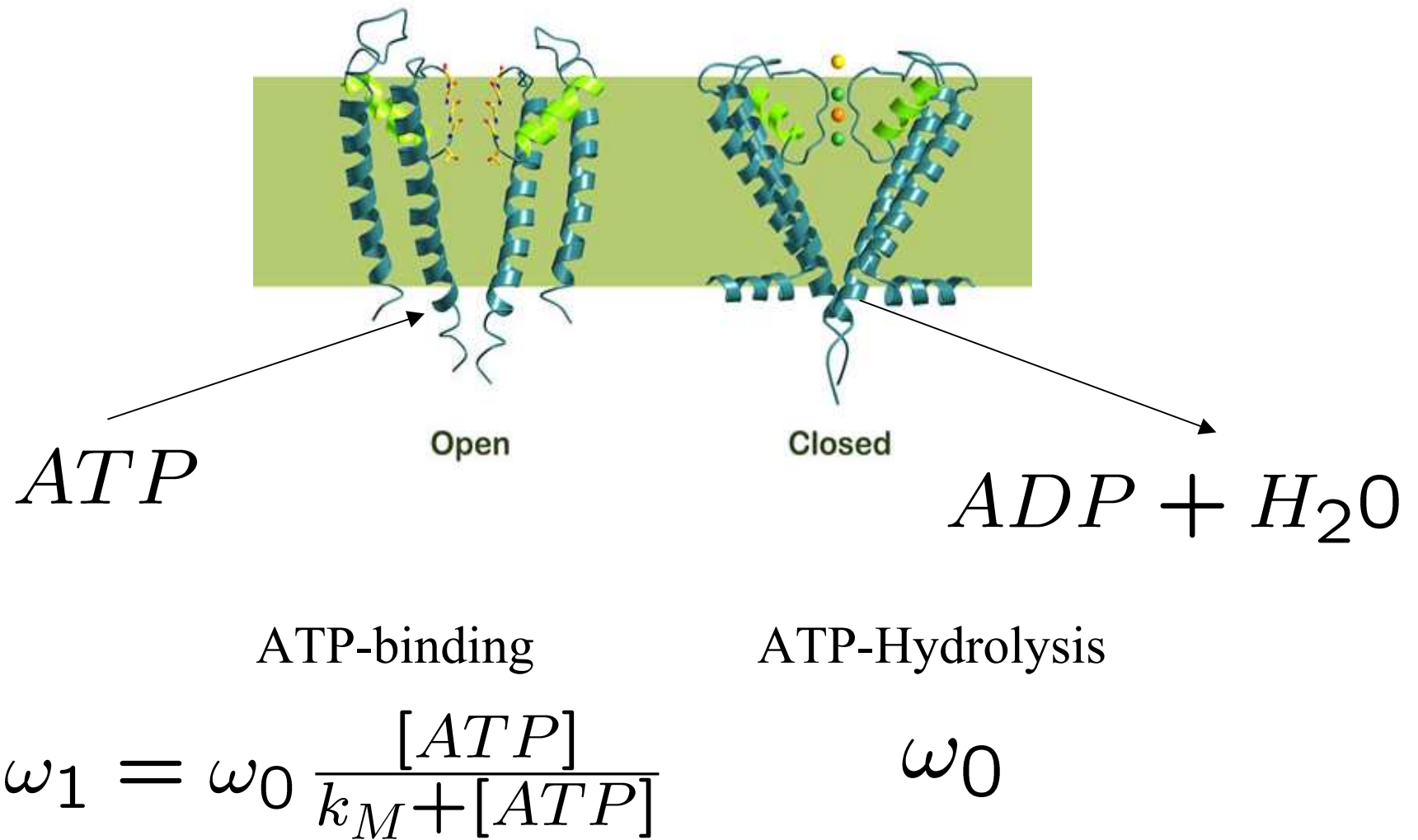
$$J = -\frac{AD}{L} (\rho_1 - \rho_0) \quad \rho_0 > \rho_1$$

$$Cl^-, \quad A = 0.2 \text{ nm}^2, \quad L = 5 \text{ nm}, \quad \Delta\rho = 0.1 \text{ M}$$

$$D = \frac{K_B T}{6\pi r_o \eta} = 1.1 \cdot 10^9 \text{ nm}^2/\text{s}$$

$$J \sim 6 \cdot 10^6 \text{ ions/s} \sim 0.4 \text{ pA}$$

ATP gated channel \rightarrow two states : open and closed.



$$J_{ATP} = -\frac{\sigma}{1+2\sigma} \frac{DA}{L} (\rho_1 - \rho_0)$$

$$\sigma = \frac{[ATP]}{k_M}$$

Free channel with potential membrane

$$J = \frac{AD}{L} \bar{v} \frac{\rho_0 - \rho_1 e^{-\bar{v}}}{1 - e^{-\bar{v}}}, \quad \bar{v} = \frac{\Delta\phi}{k_B T}$$

Goldman-Hodkin-Katz eq.

Voltage gated channel: Na

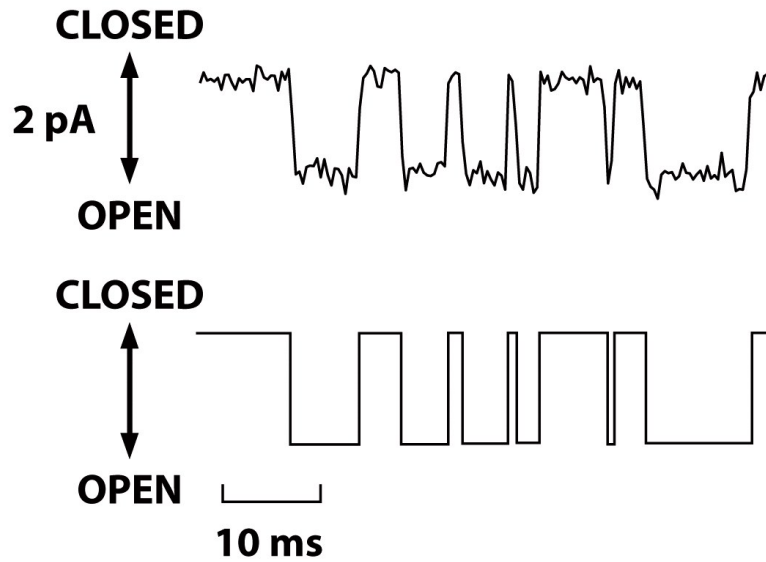


Figure 7.2a Physical Biology of the Cell (© Garland Science 2009)

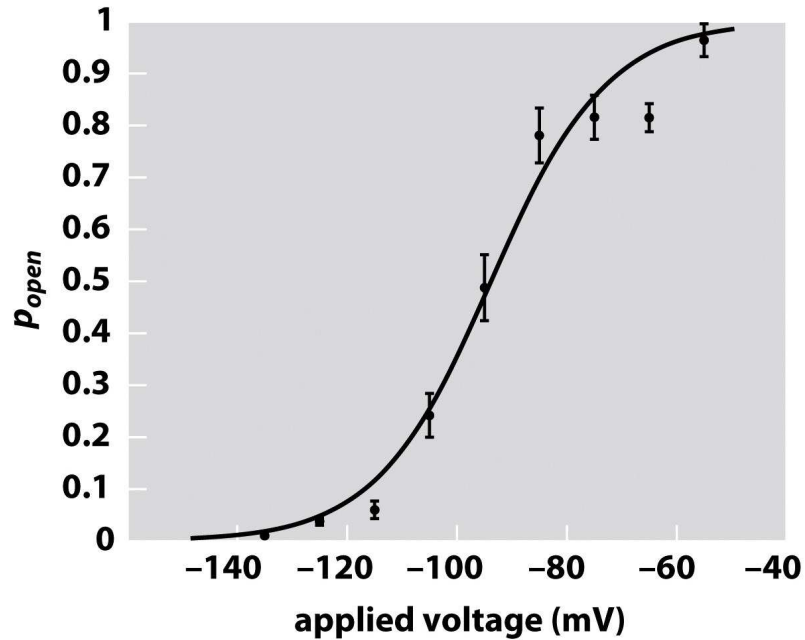
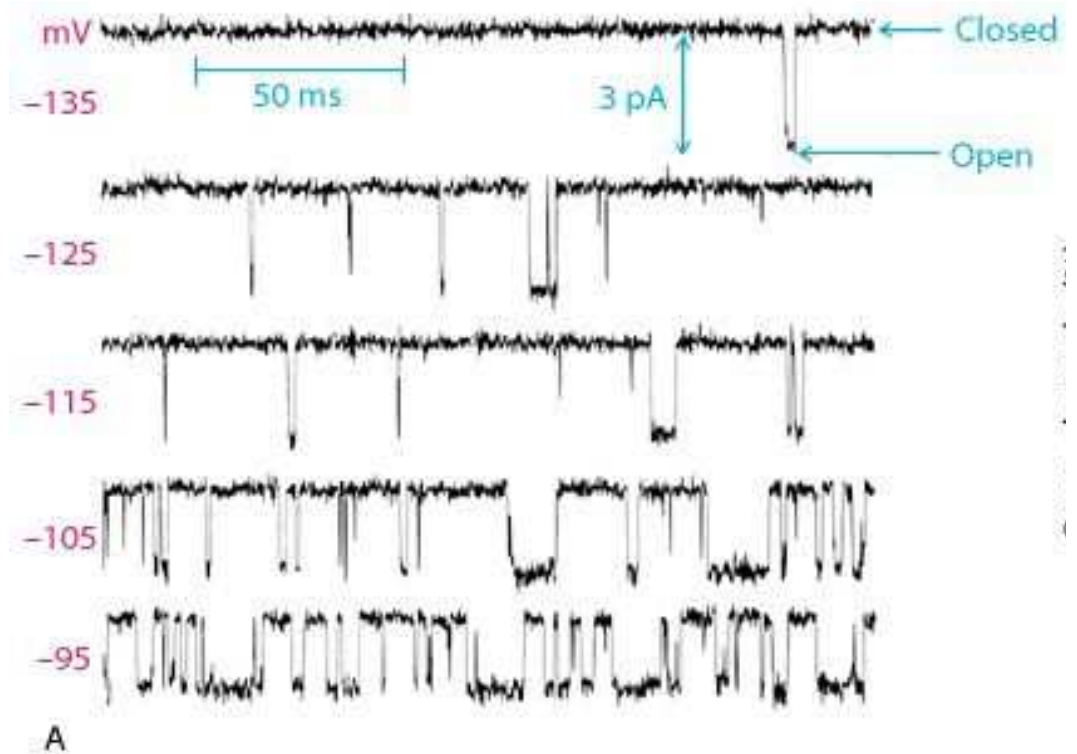


Figure 7.2c Physical Biology of the Cell (© Garland Science 2009)

Na reconstructed channels,
from Hartshorne et al. PNAS **82**,240(1985)



PUMPS

Particle flux is against gradient

power in > power out

$$\frac{\Delta G_{ATP}}{t_{ATP}} > J\mu$$

$$\frac{\rho_1}{\rho_0} = 10 \rightarrow \mu = k_B T \ln 10 = 2.3 k_B T / \text{particle}$$

$$t_{ATP} = 1 \text{ms}$$

$$J < 10^3 \text{ ions/s} = 0.16 \cdot 10^{-3} \text{ pA}$$

WHAT ELSE ?

Put a ratchet in your model!

Overdamped Langevin equation

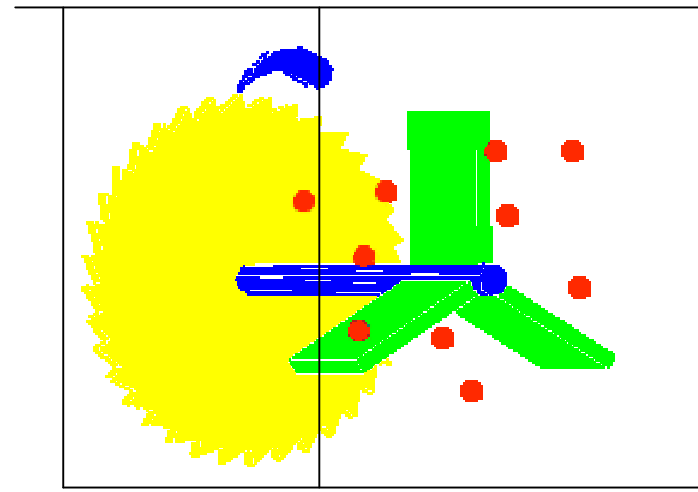
Two states ratchet potential

ATP dependent transitions

Energy constrains



Feynman, 1963

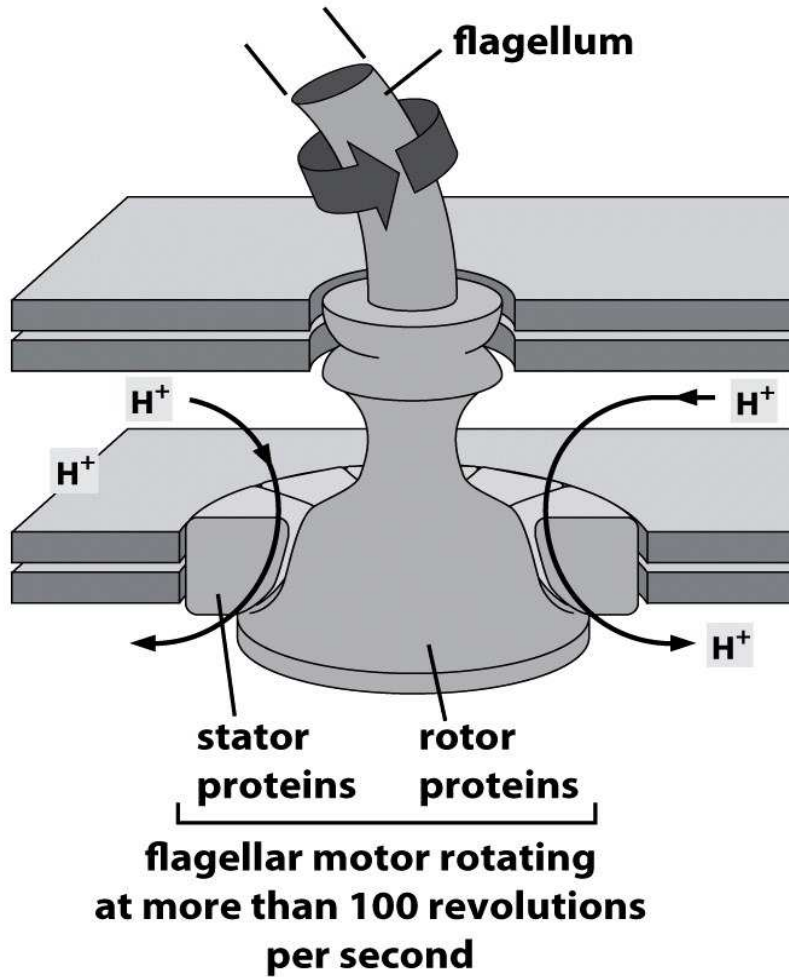


$$T_1 < T_2$$

2. MOTORS

Bacterial Flagellar Motor (BFM)

Chien-Jung, et al. Bioph. J. 93, 294 (2007)

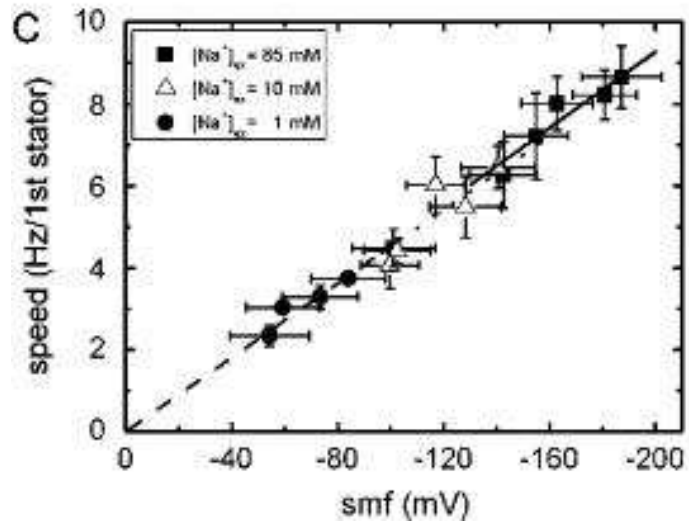


$$smf = -\Delta\phi + k_B T \ln \frac{[Na]_{in}}{[Na]_{ex}}$$

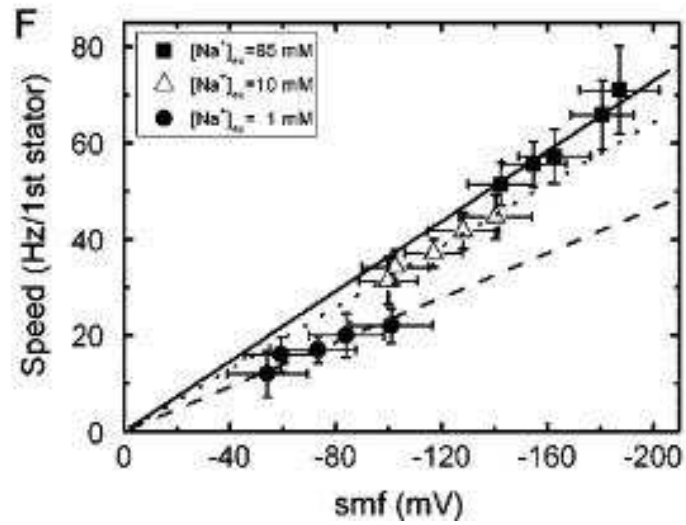
$$\omega = \frac{1}{t_0 \left(1 + \frac{k}{[Na]} \right) + \frac{\gamma_0 + \gamma_L}{-smf}}$$

Figure 16.13a Physical Biology of the Cell (© Garland Science 2009)

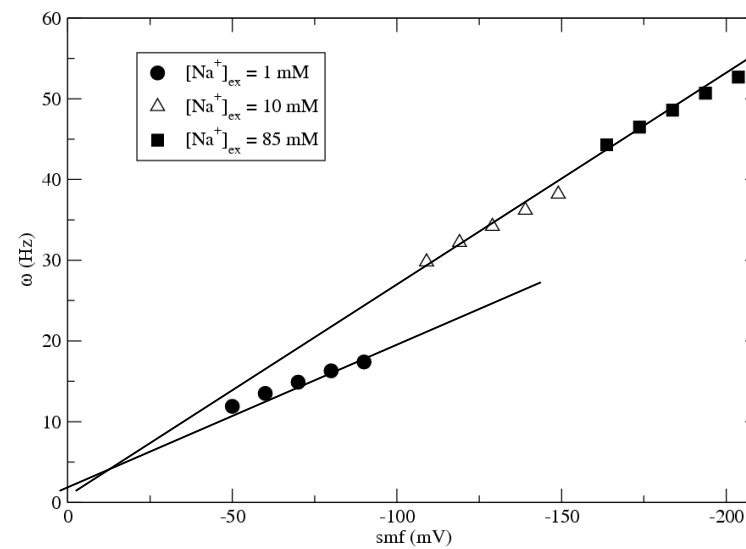
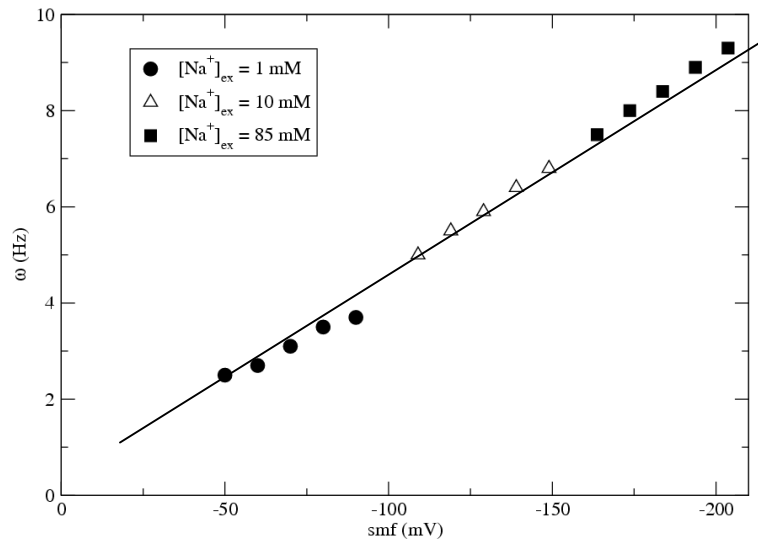
Experiment $1\mu m$



Experiment $0.35\mu m$

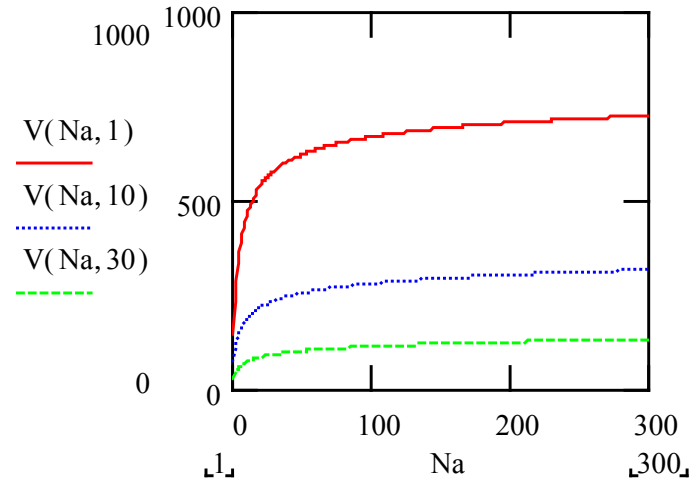
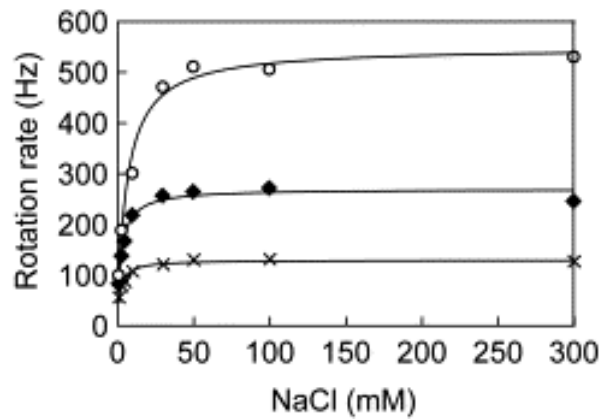


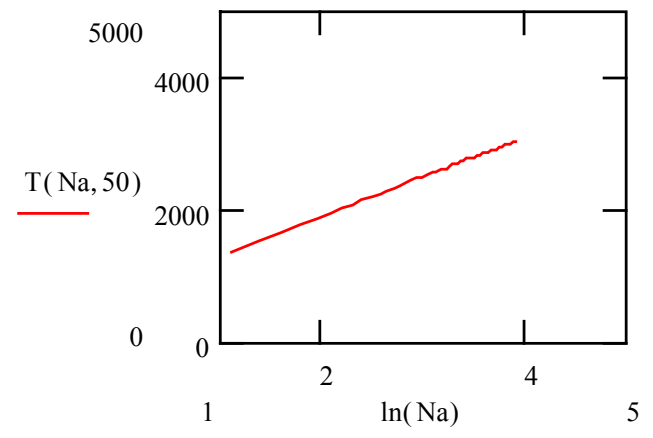
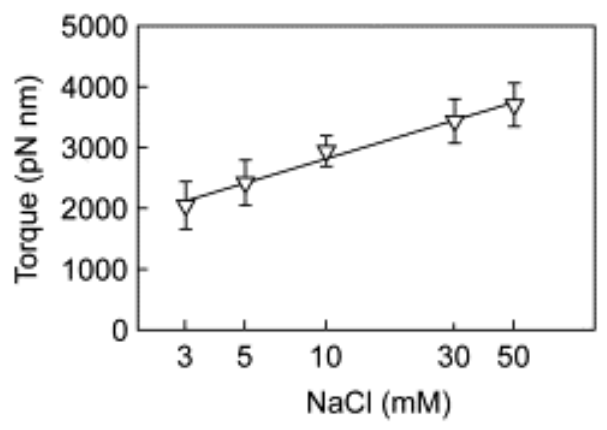
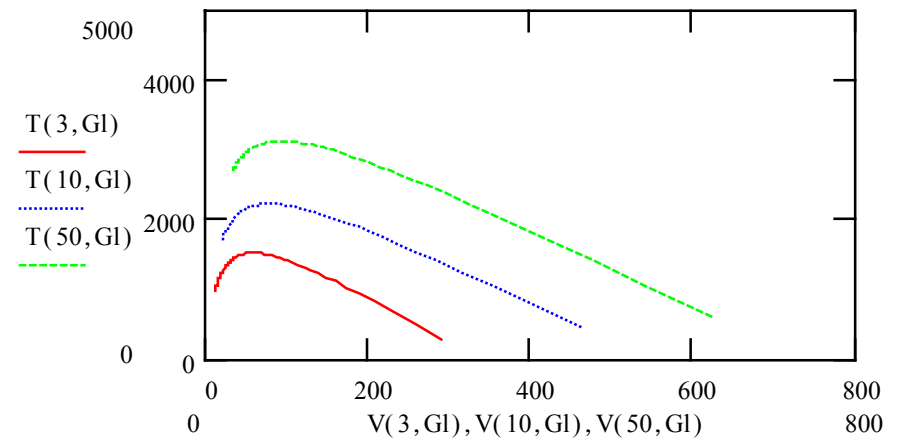
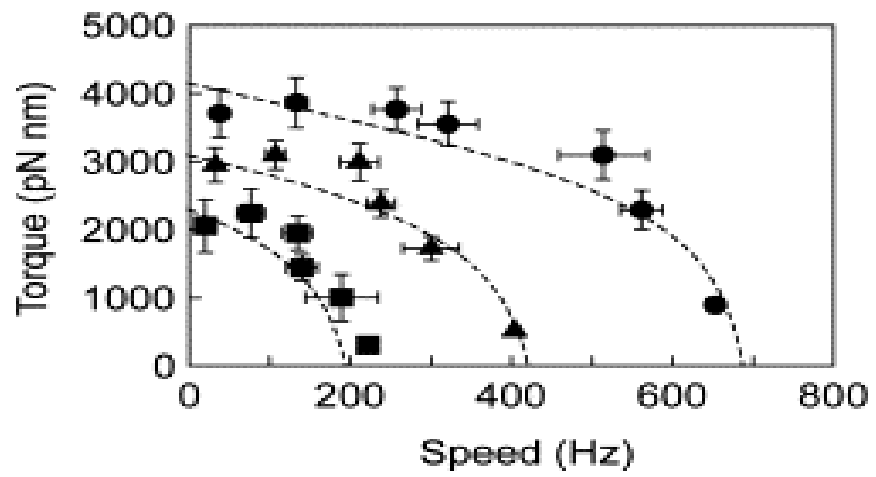
Theory



Sowa et al, JMB 327, 1043(2003)

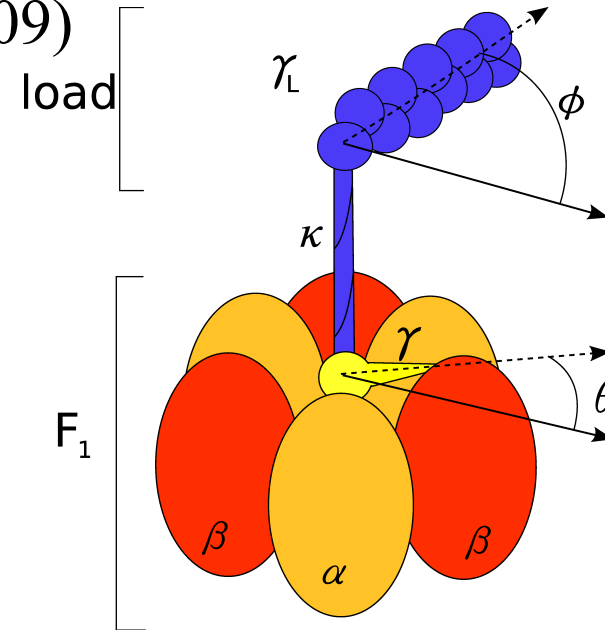
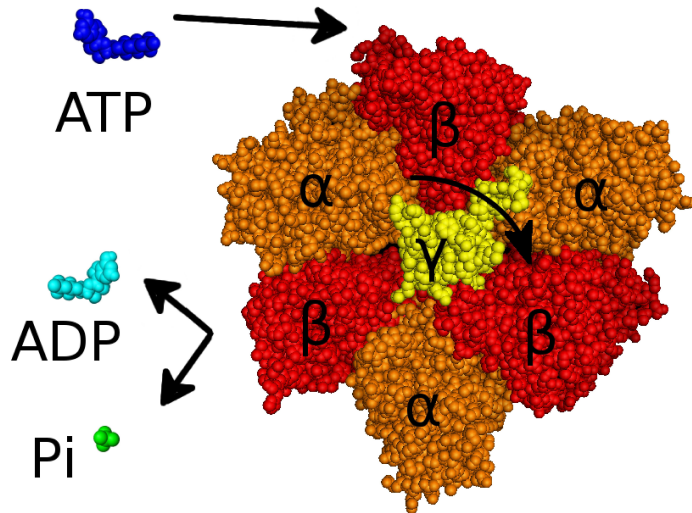
$$\omega = \frac{L_0}{t_0 \left(1 + \frac{k}{[Na]} \right) + \frac{(\gamma_0 + \gamma_L)L_0^2}{-smf - b * R}}$$





F1-ATPase

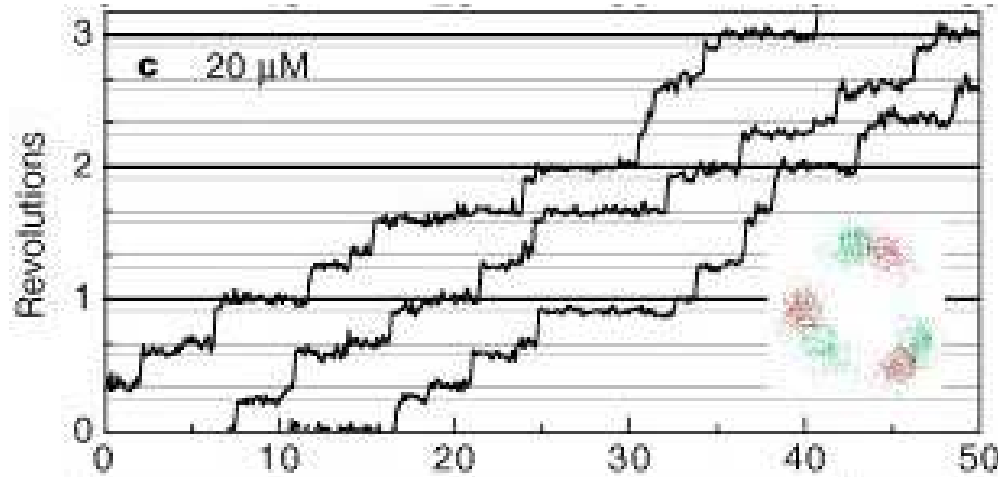
Theoretical analysis of the F1-ATPase experimental data,
R. Perez-Carrasco & JMS, preprint (2009)



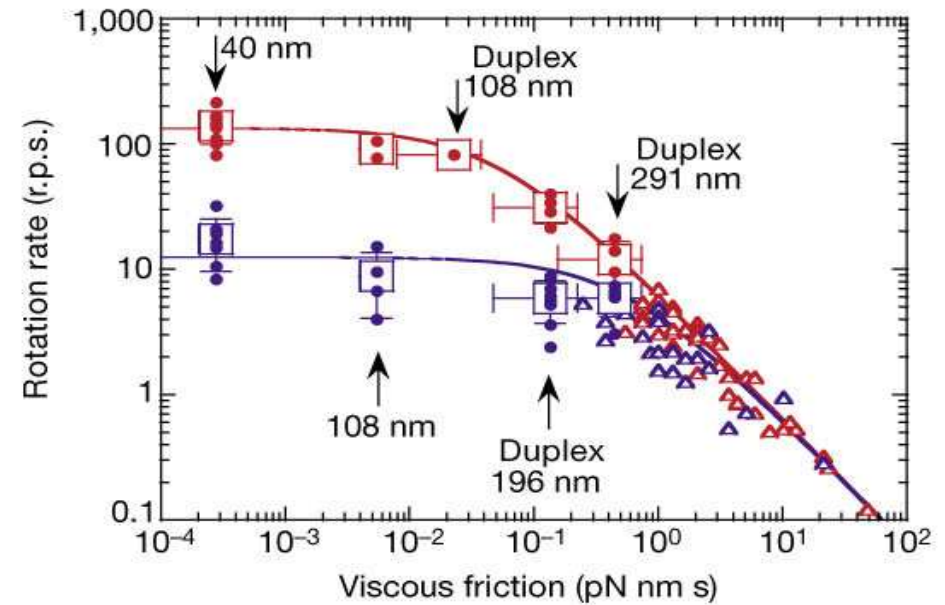
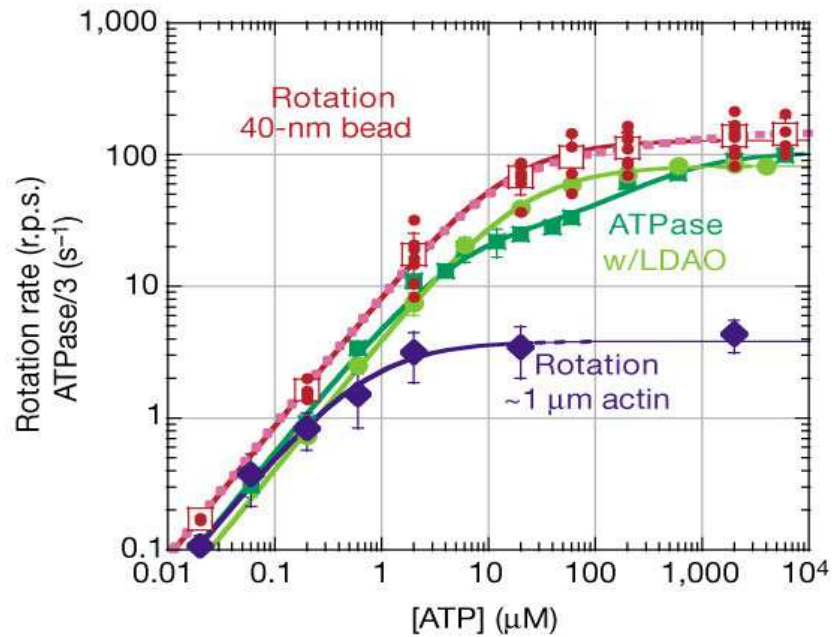
Observable: load angle $\phi(t) \rightarrow \omega$

Parameters: load friction γ_L and $[ATP]$

Experimental data from: Yasuda et al., Nature **410**, 898 (2001)

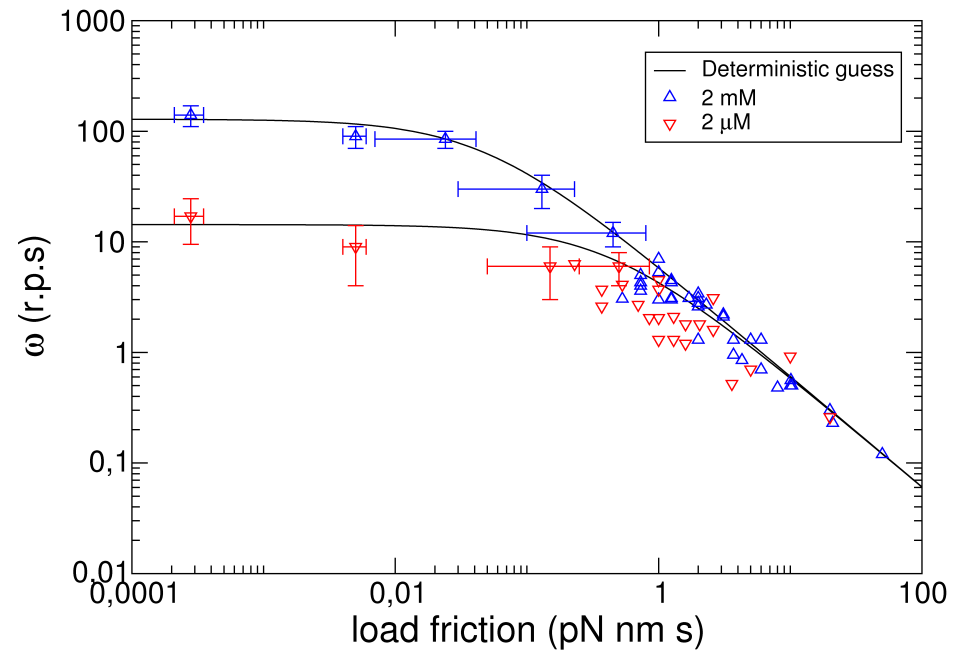
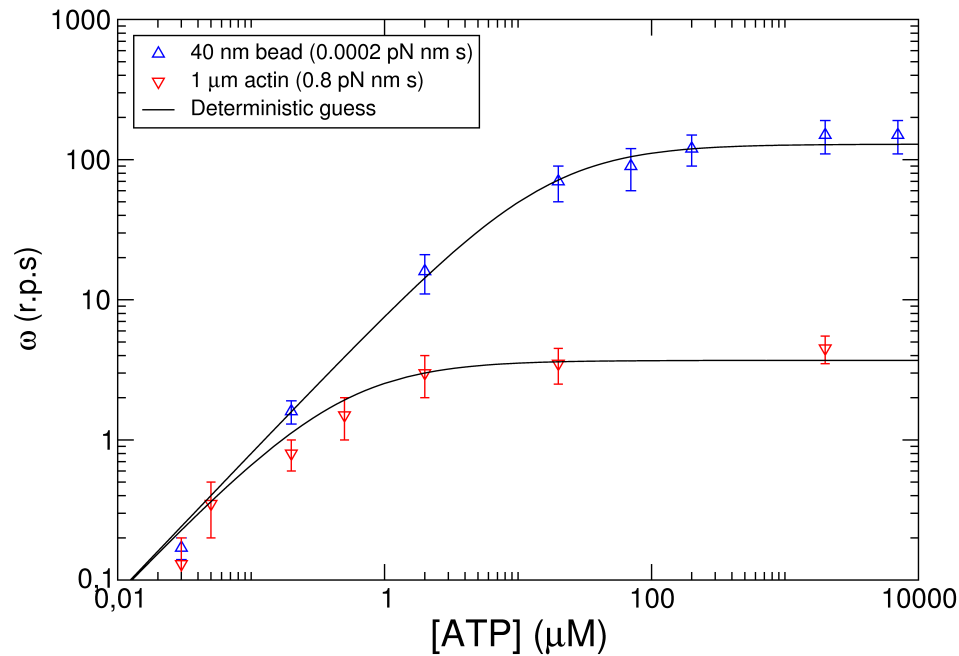


Substeps: $90^\circ + 30^\circ$



First approach

$$\omega = \frac{\theta_0}{t_0 \left(1 + \frac{k}{[ATP]} \right) + \frac{\theta_0^2 (\gamma_0 + \gamma_L)}{\Delta G_{ATP}}}$$



Parameters: V_0 , E_1 , E_2 , and α .

Conditions:

$$E_1 = V_0 \left(1 - \frac{1}{3\alpha}\right)$$
$$E_2 = 3V_0(1 - \alpha)$$
$$E_1 + E_2 = \Delta G_{ATP}$$
$$\alpha$$

Optimization: ω small \rightarrow t maximum

$$\alpha = 0.805 \rightarrow \theta_1 = 84.85^\circ \quad \theta_2 = 35.15^\circ$$

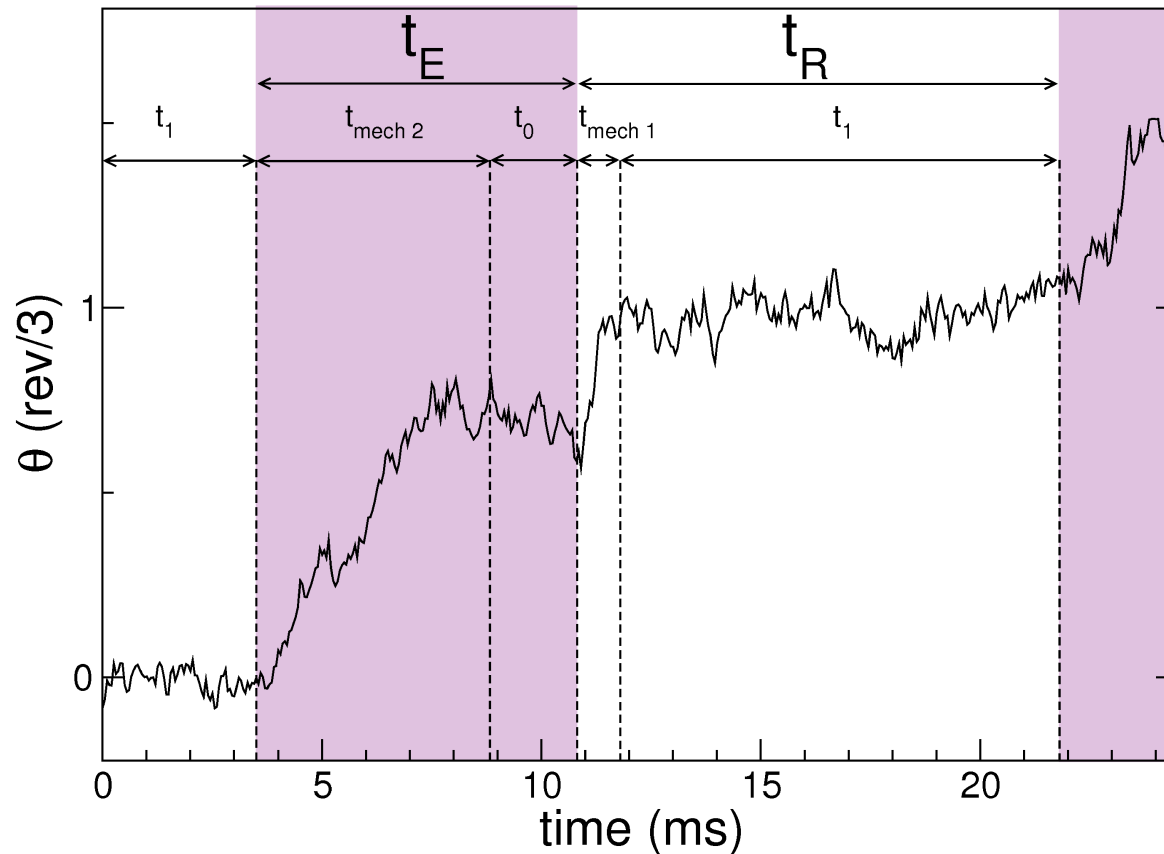
Torques

$$\tau_1 = 27 \text{ pN.nm}, \tau_2 = 65 \text{ pN.nm}, \bar{\tau} = 40 \text{ pN.nm}$$

Dynamical equations

$$\gamma \dot{\theta} = -V(\theta, [ATP]) + k(\phi - \theta) + \xi(t)$$

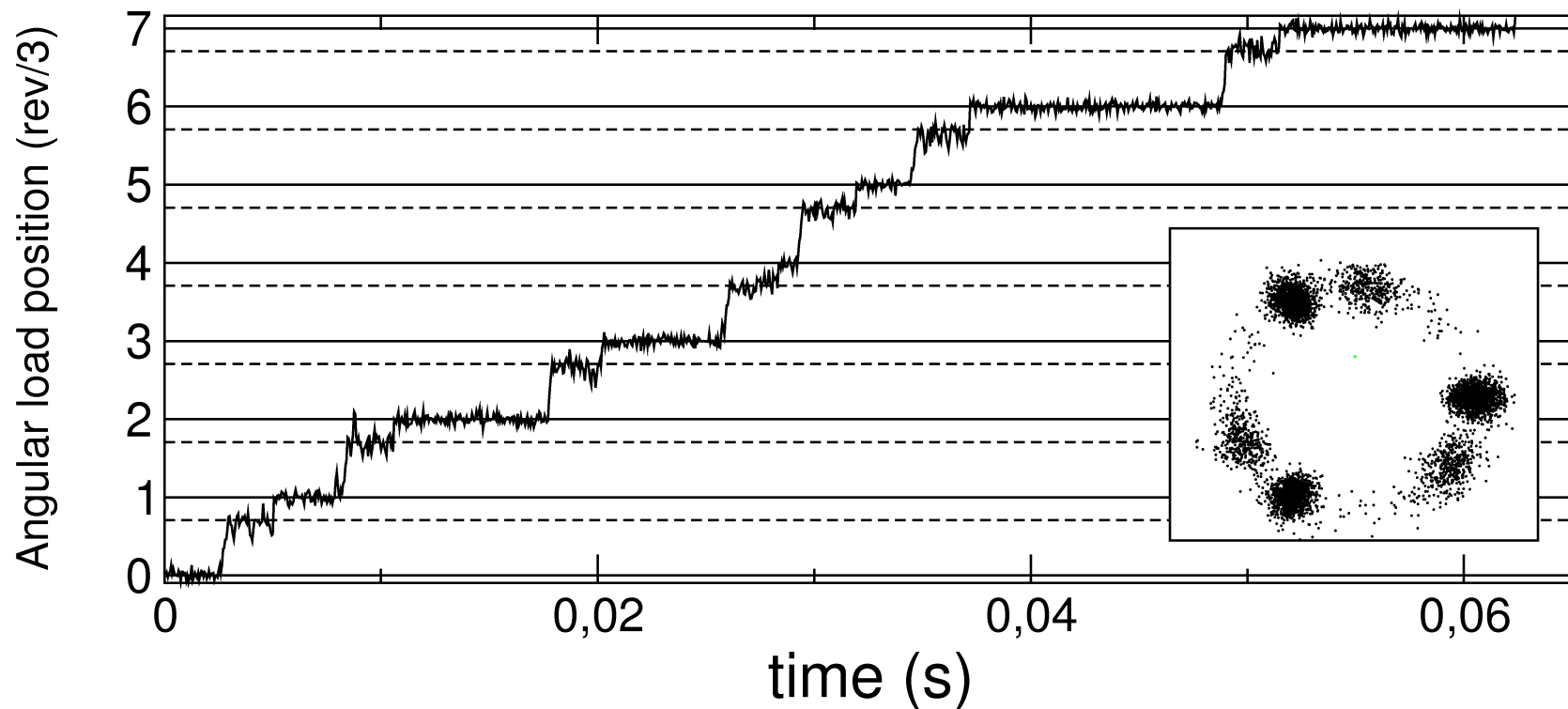
$$\gamma_L \dot{\phi} = -k(\phi - \theta) + \xi_L(t)$$

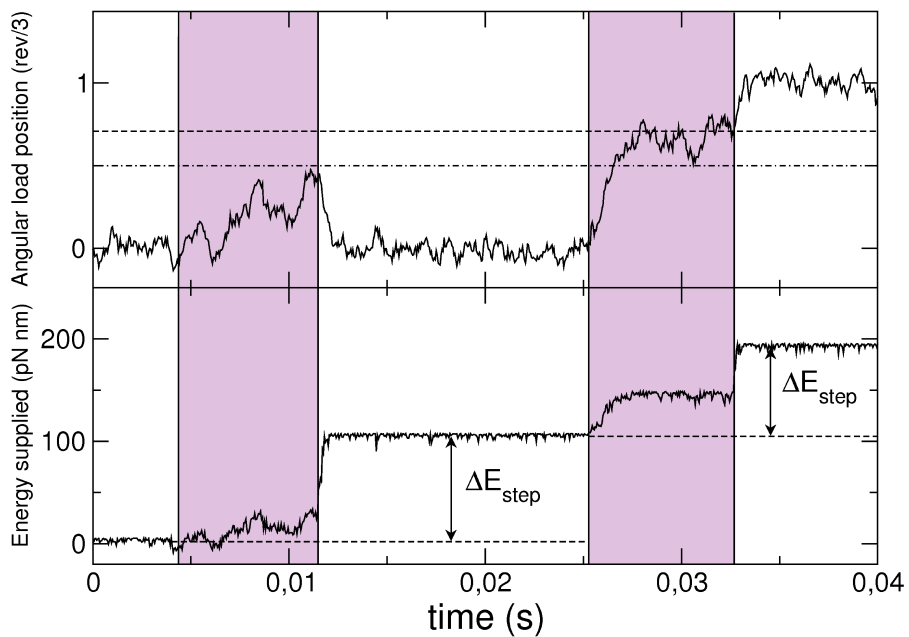
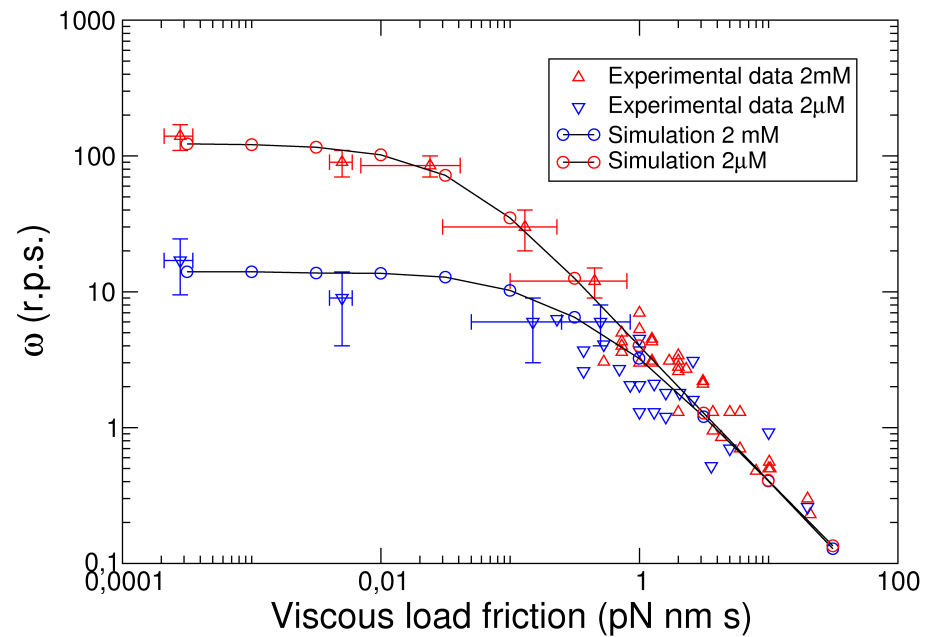
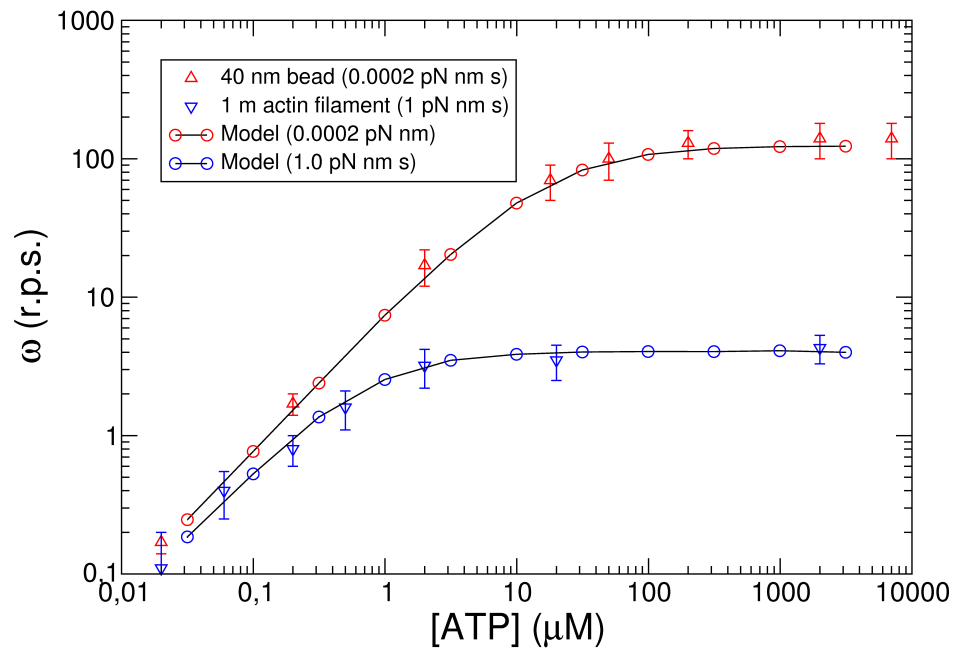


$$t_R = t_{mech1} + t_1, \quad t_E = t_{mech2} + t_0$$

$$P(t_1) = \frac{1}{t_{ATP}} e^{-t_1/t_{ATP}}$$

$$t_{ATP} = t_0 \frac{k}{[ATP]}$$





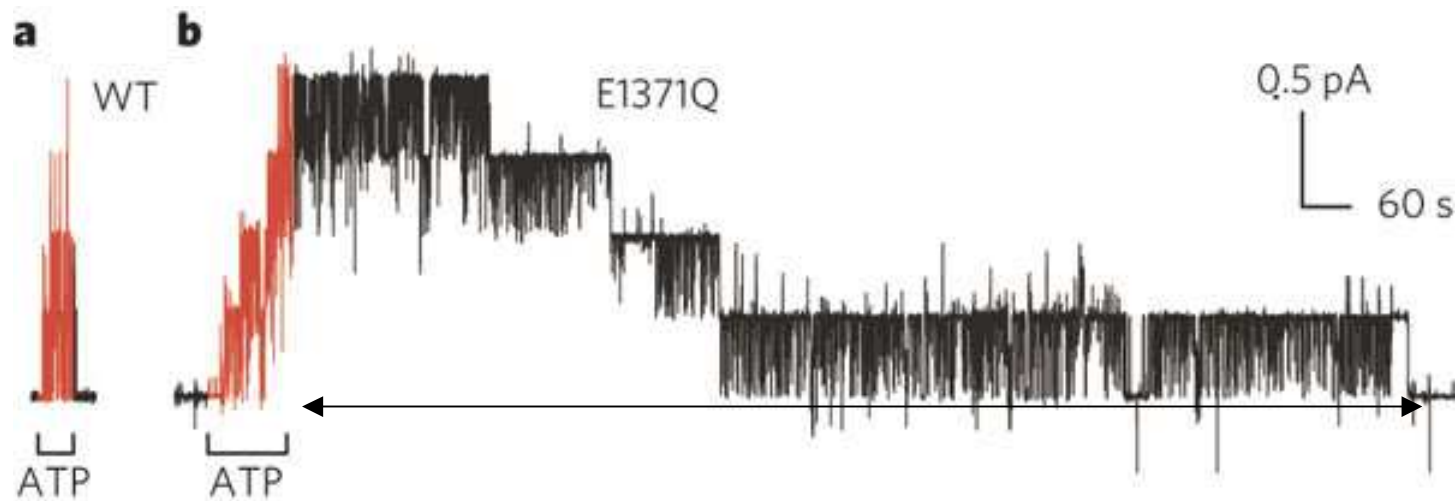
Input energy
 coupling ratio

3. CHANNELS

CFTR chloride channel (cystic fibrosis)

Gadsby, et al, NATURE 440,477(2006)

Experimental data: channels need energy input



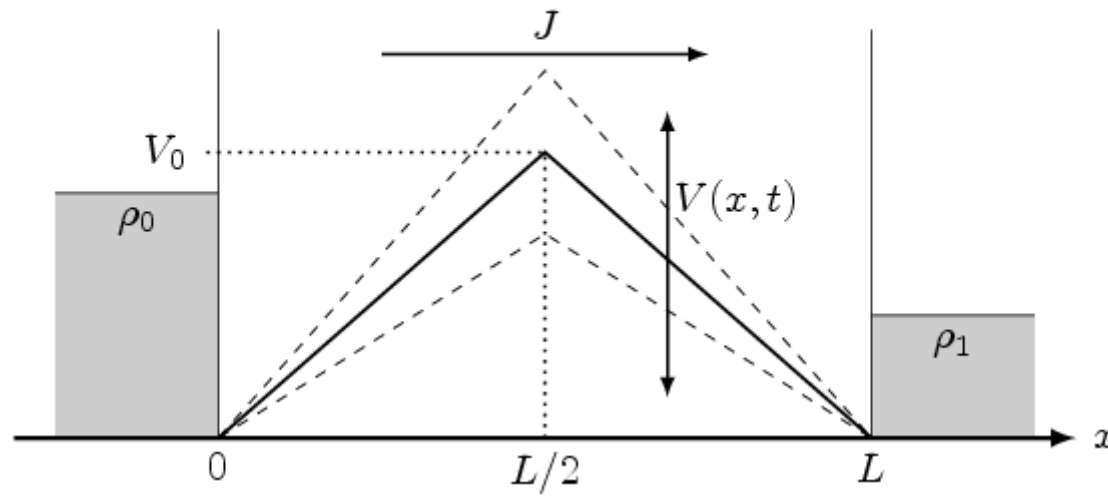
ATP-binding

OPEN

ATP-hydrolysis

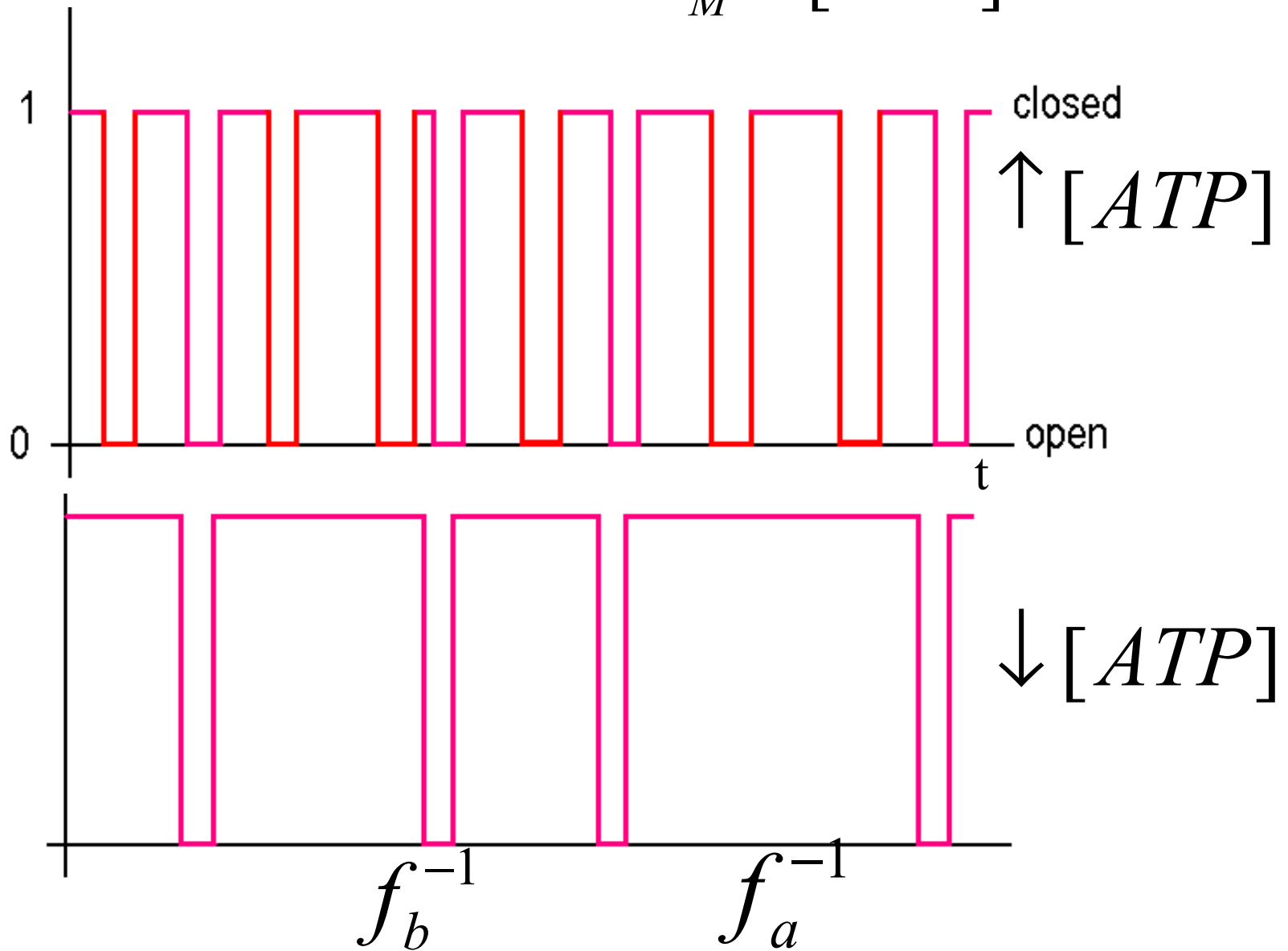
CLOSED

Simple model

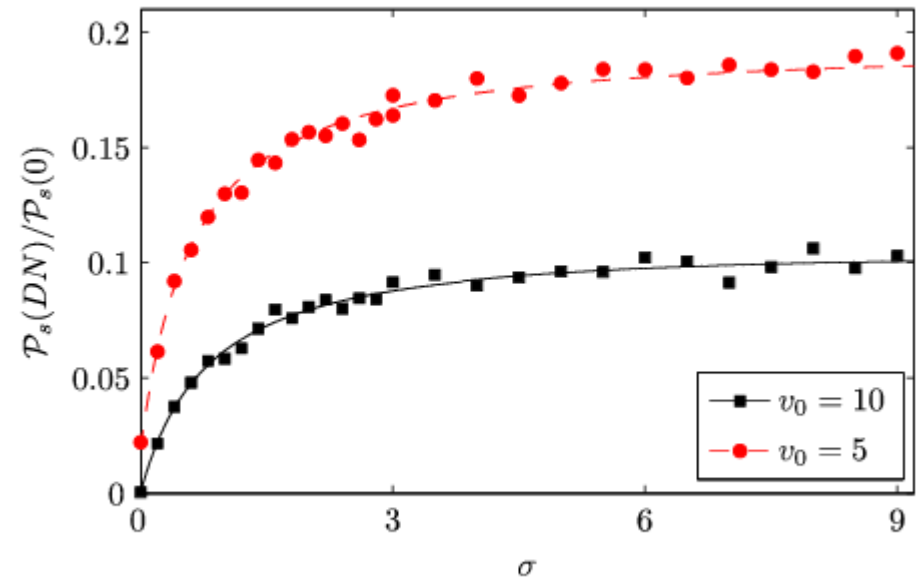
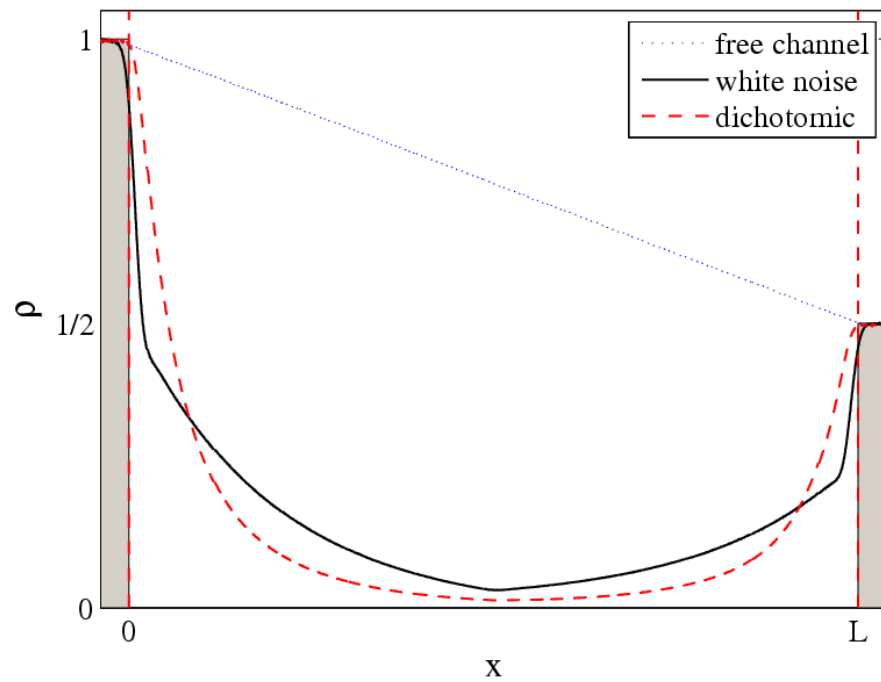


$$\gamma \dot{x} = -V(x, [ATP]) + \xi(t)$$

$$f_b = \omega_0 \qquad f_a = \frac{\omega_0 [ATP]}{k_M + [ATP]}$$

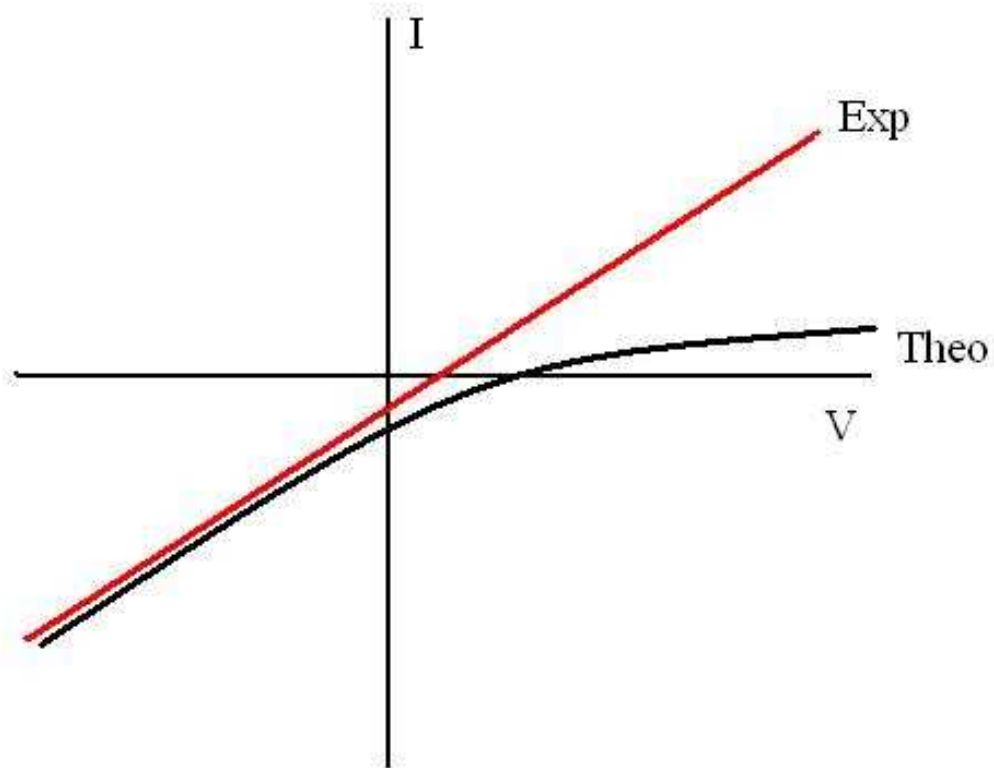


Theoretical study of a membrane channel gated by ATP
J.G. Orlandi & JMS, Eur.Phys.J. (2009)



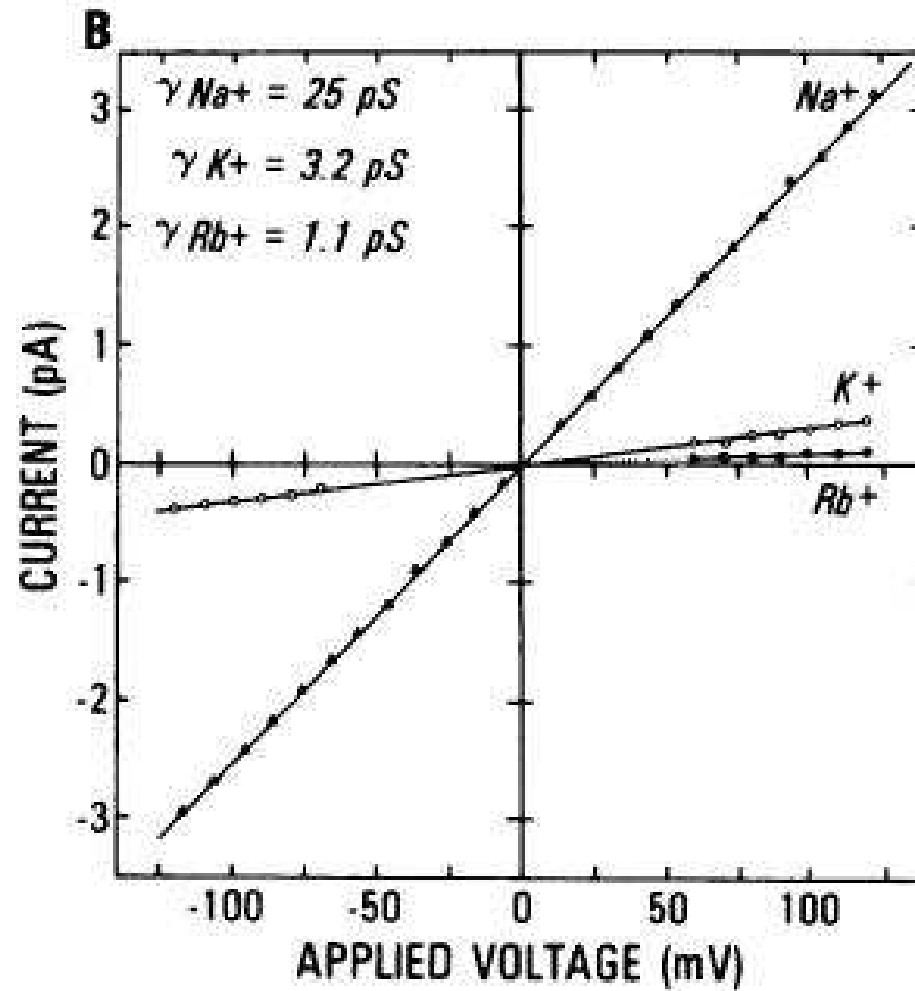
...but...experimental data for CLC Cl channel...

Accardi et al. J. Gen. Physiol., 123, 109(2004)



Improvement ? : structured potential inside the channel

Selectivity

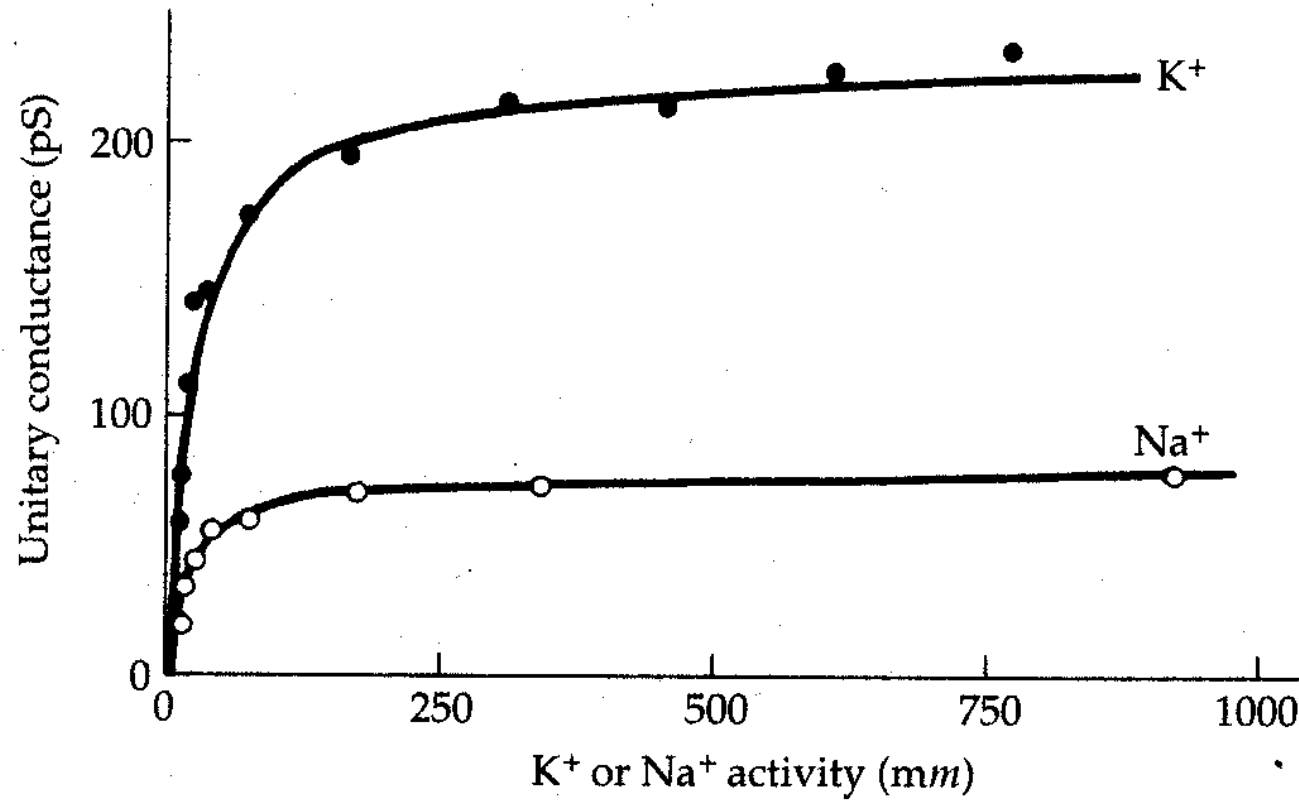


from Hartshorne et al. PNAS **82**,240(1985)

SATURATION

$$J = \frac{J_{max}[S]}{K_s + [S]}$$

(B) SARCOPLASMIC RETICULUM CHANNEL



Na anomalous conductance?

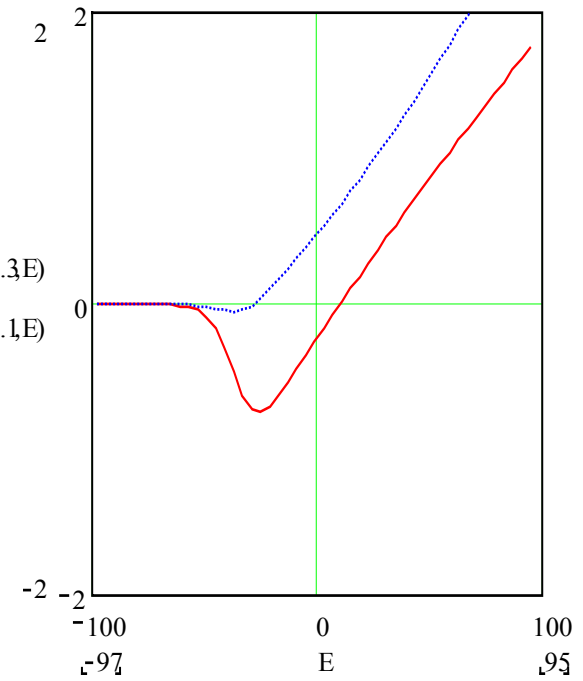
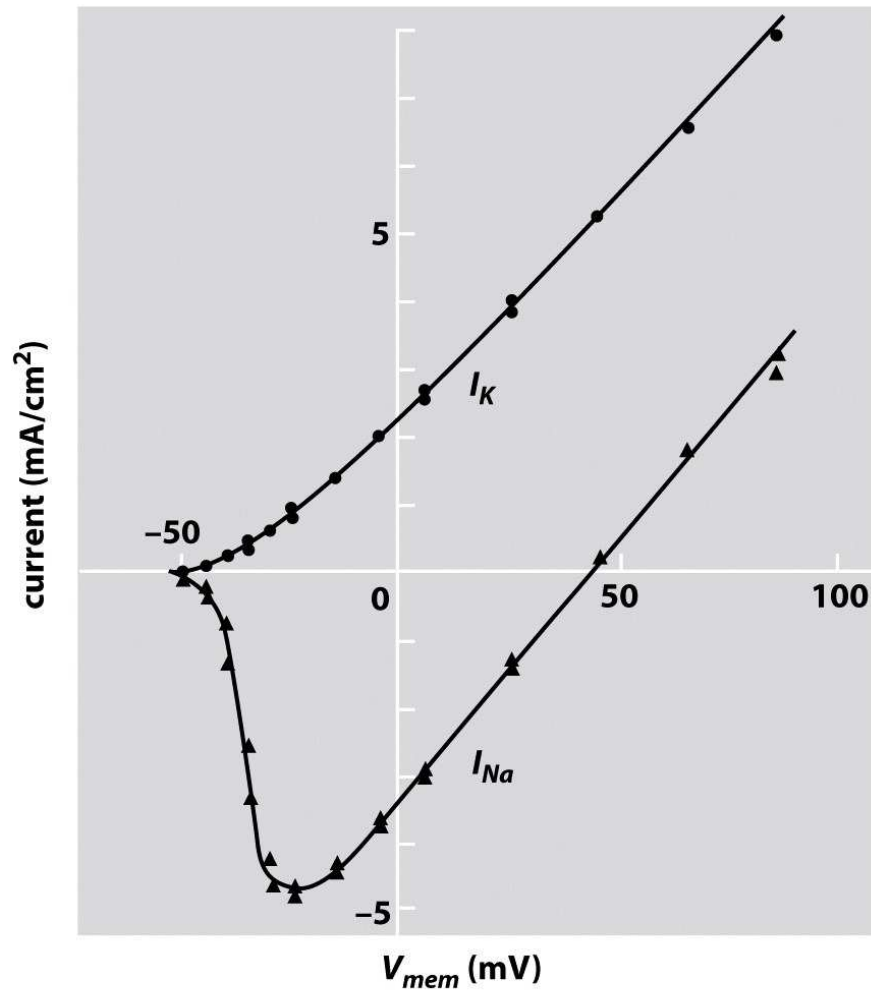
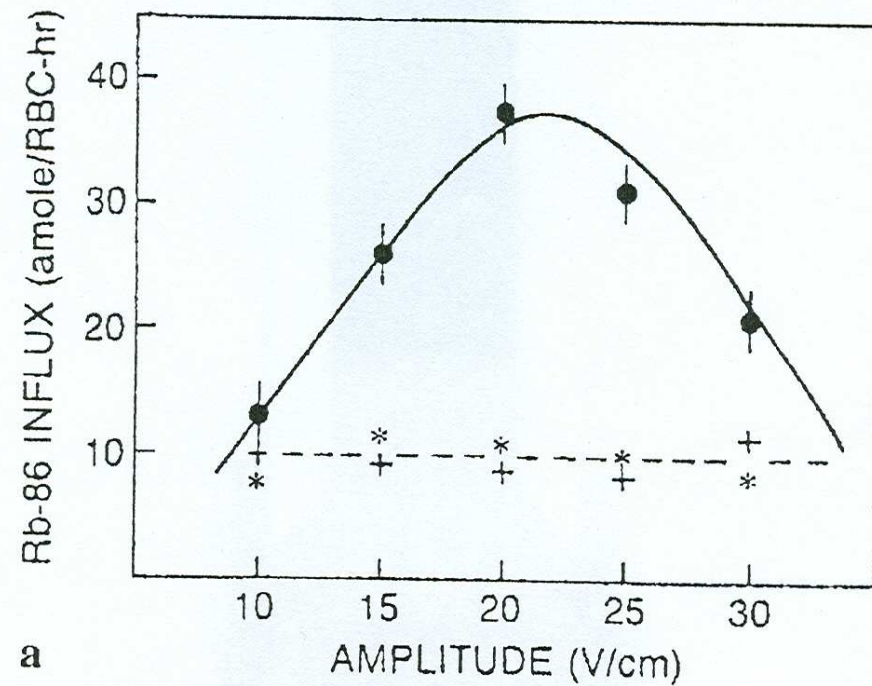
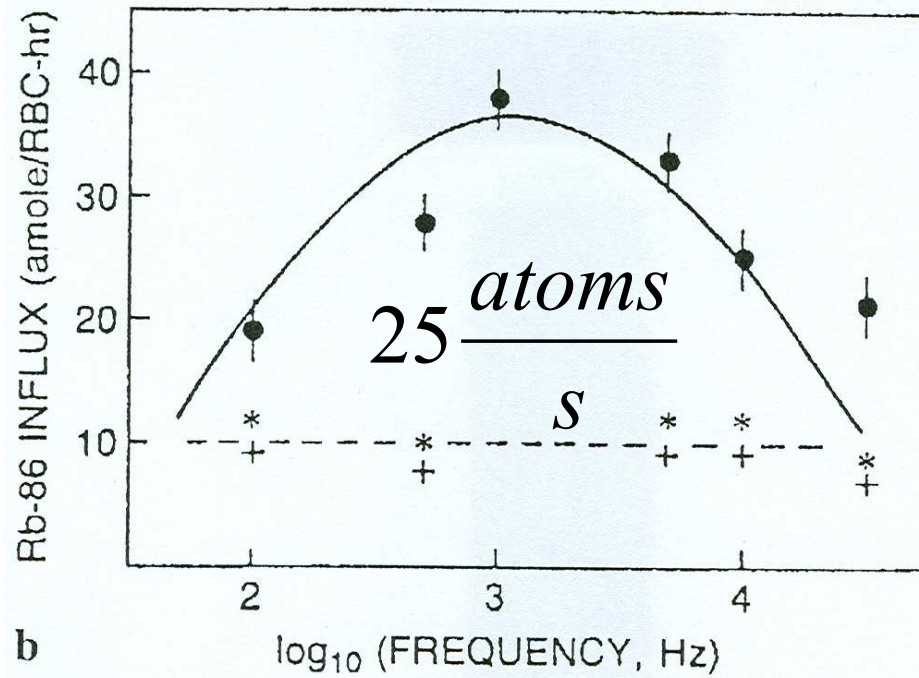


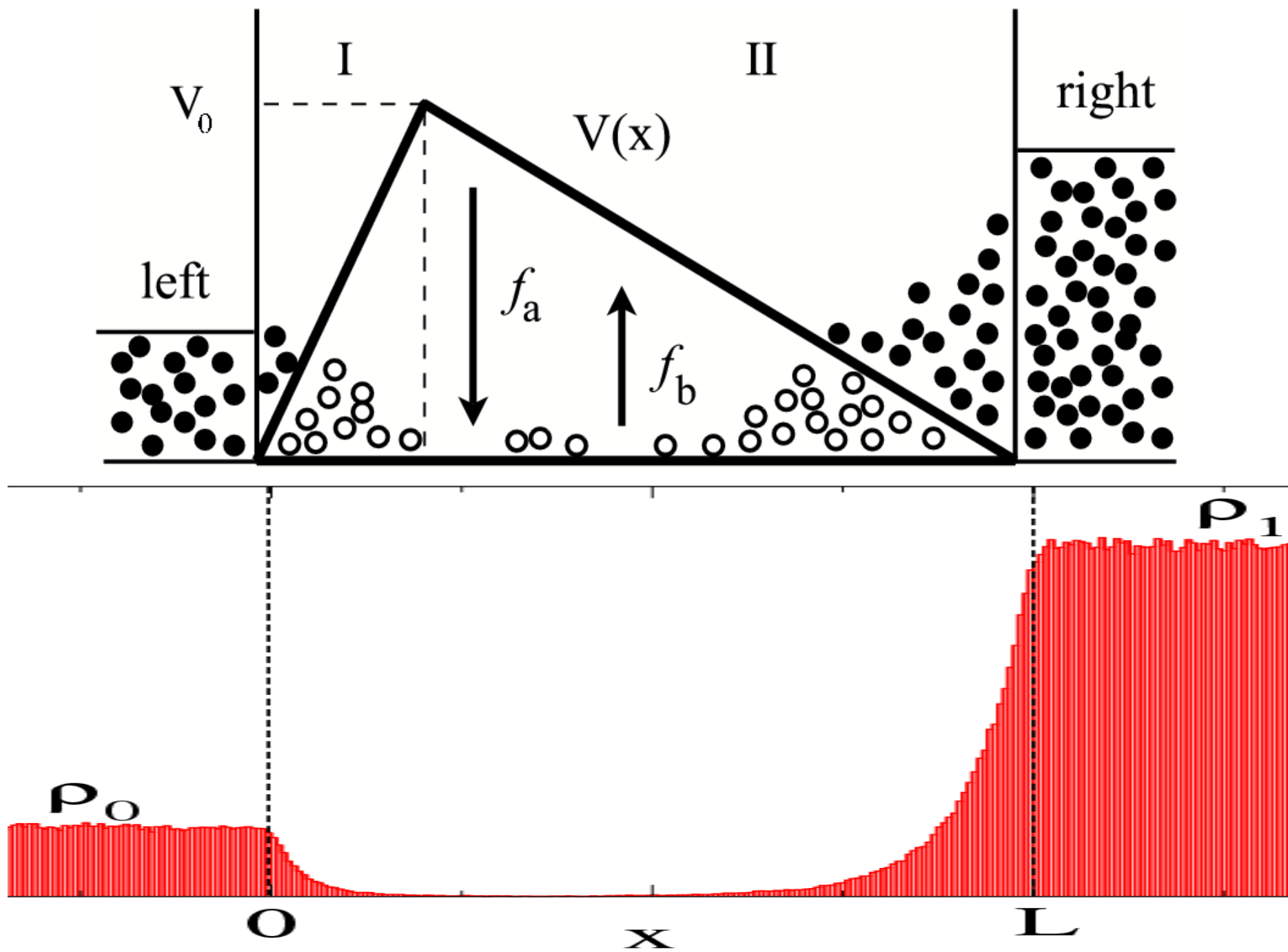
Figure 17.12 Physical Biology of the Cell (© Garland Science 2009)

4. PUMPS

Na-K pump

Tsong & Xie, Appl.Phys. A75,345(2002)





$$\left[d_z^3 - 2F_i d_z^2 - (f_a + f_b) d_z + 2f_b F_i \right] \rho_i(z) = J(f_a + f_b)$$

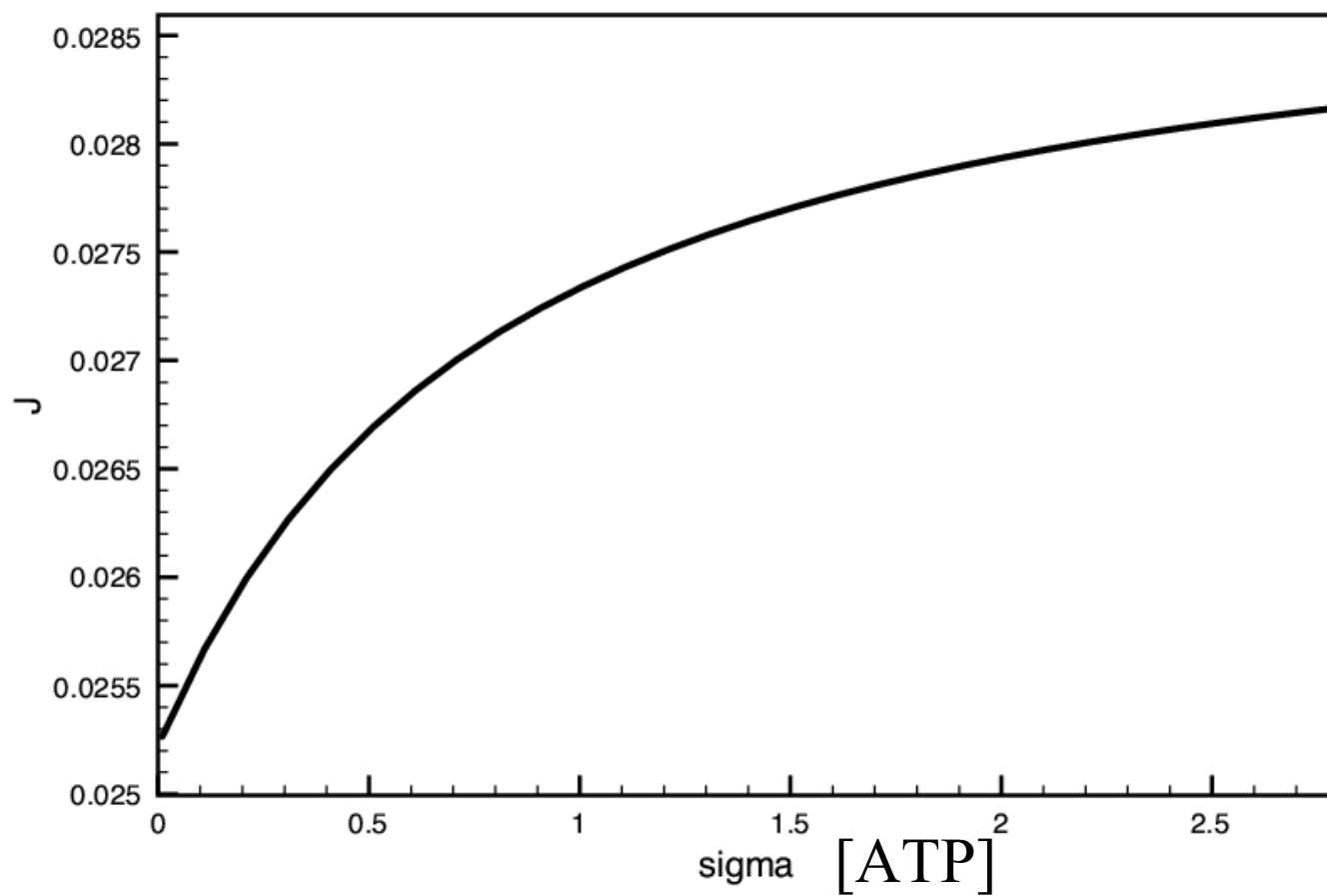
Seven unknown constants with seven boundary conditions

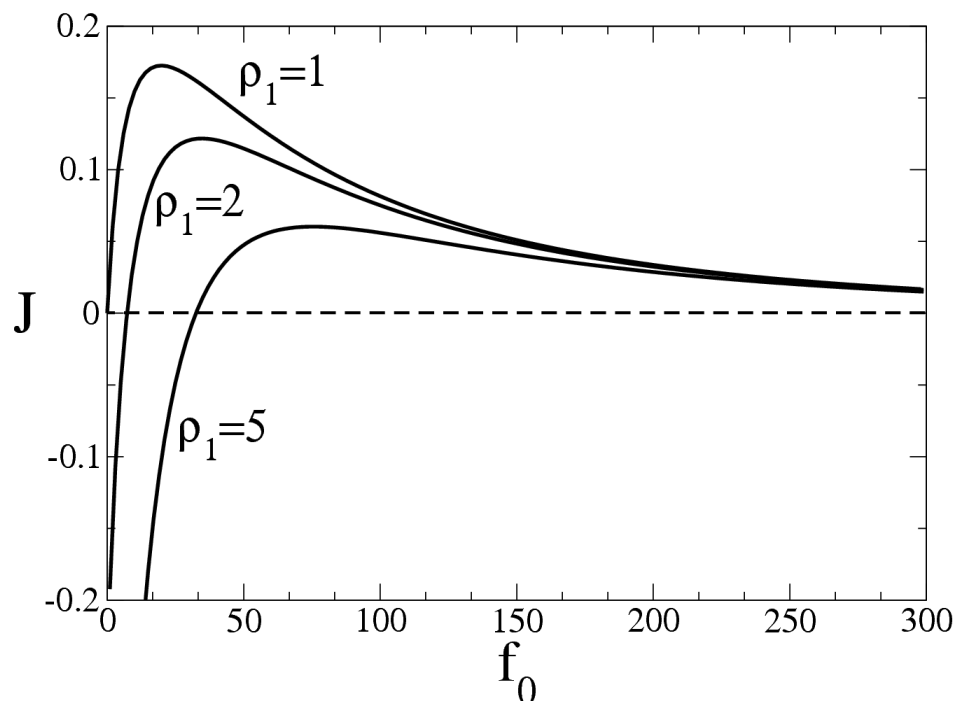
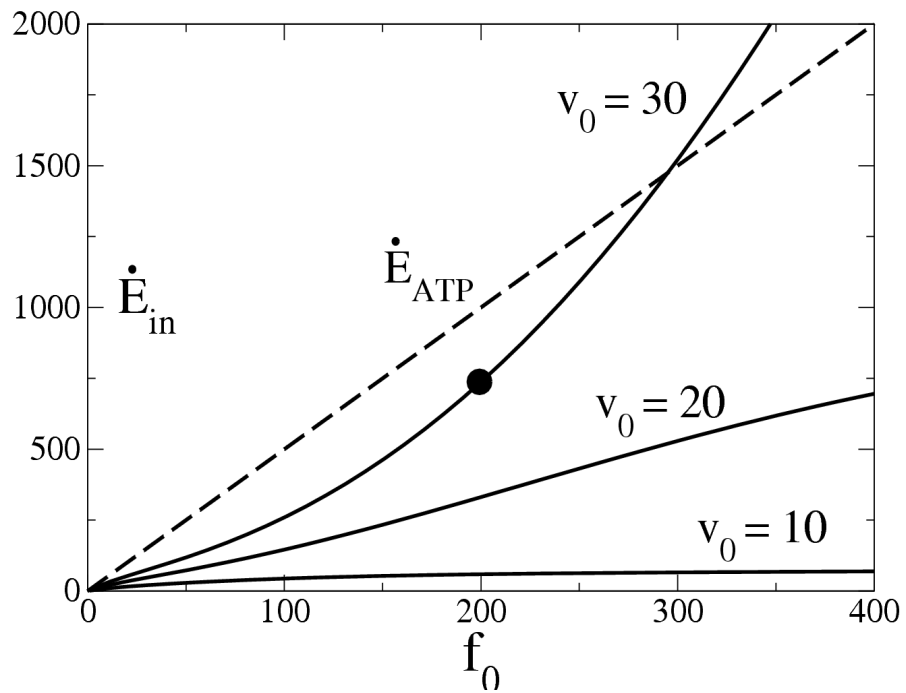
$$\text{if } J = 0 \rightarrow \frac{\rho_{\max}}{\rho_0}$$

$$\text{If } \rho_1 < \rho_{\max} \rightarrow J \neq 0$$

Parameters: $V_0, \delta, L, S, \gamma, \rho_0, \omega_0, \sigma([ATP]), \dots$

$$\omega_0 = 300, \frac{\rho_1}{\rho_0} = 10$$





2. CONCLUSSIONS

Work in progress

V gated channels

Binding sites

Selectivity

Saturation against substrate (M-M)

Rotatory pumps (mechanical model)

Motors fueled by gradient and... more: ΔG_{ATP} , τ_{ext}

Important points

-Complex systems

-Different levels of description

-Variety of models

-Experimental data available but not enough

-Biological parameter values

THANKS



Muneyuki et al, Bioph.J. 92, 1806 (2007)

